

# Proteins?

Protein function

Protein folding

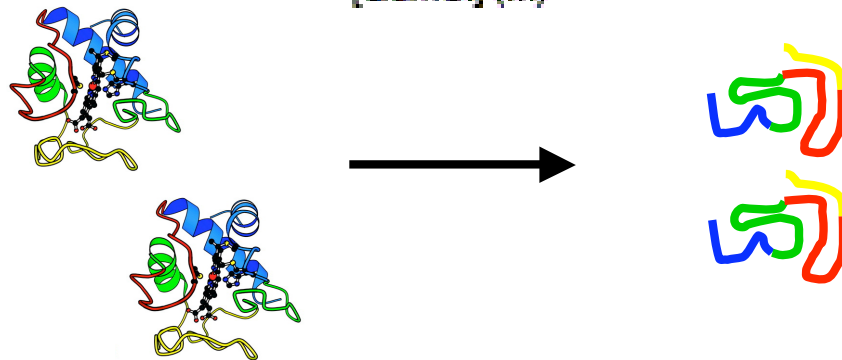
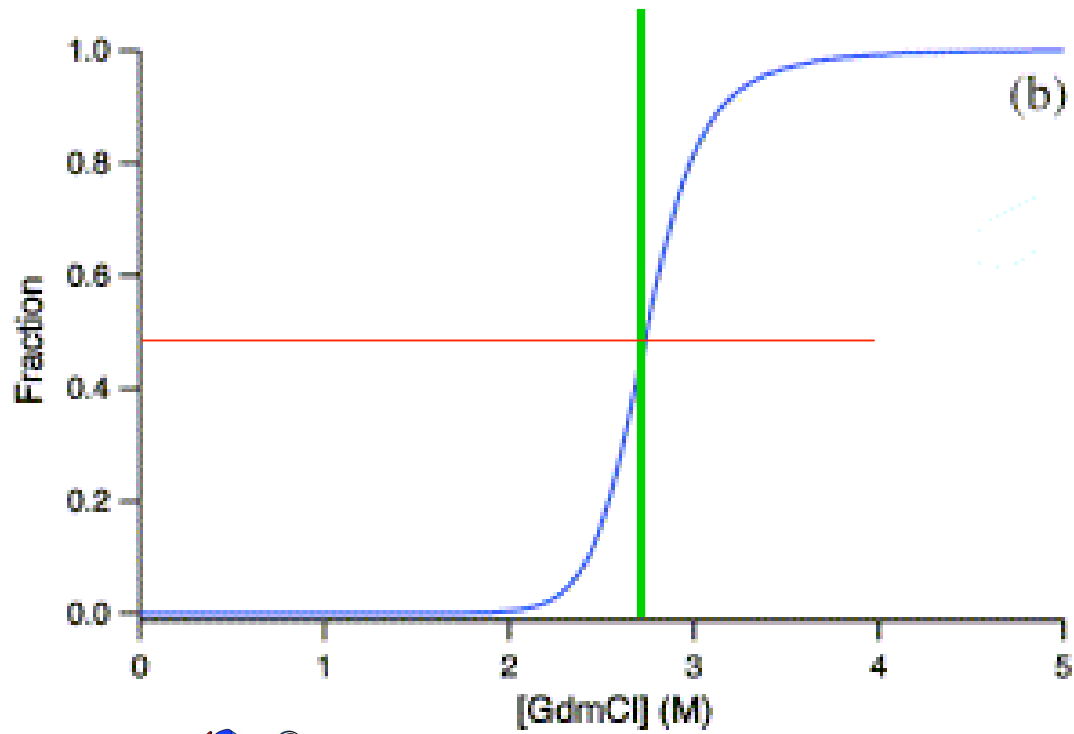
Protein folding diseases

Protein interactions

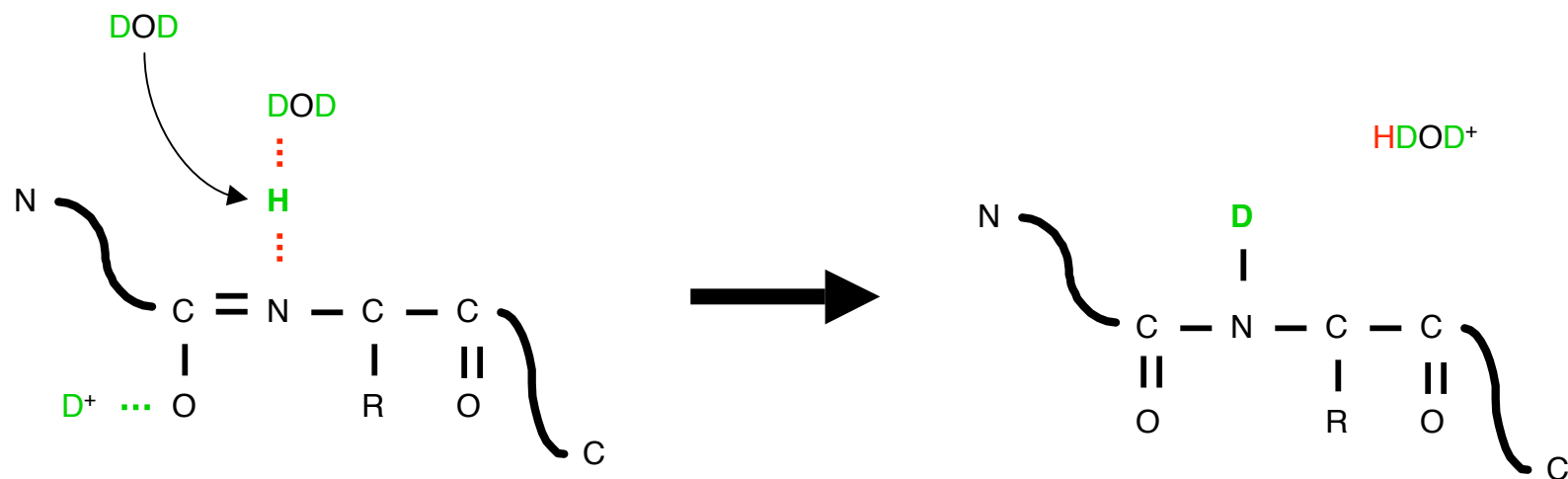
Macromolecular assemblies

The end product of Genes

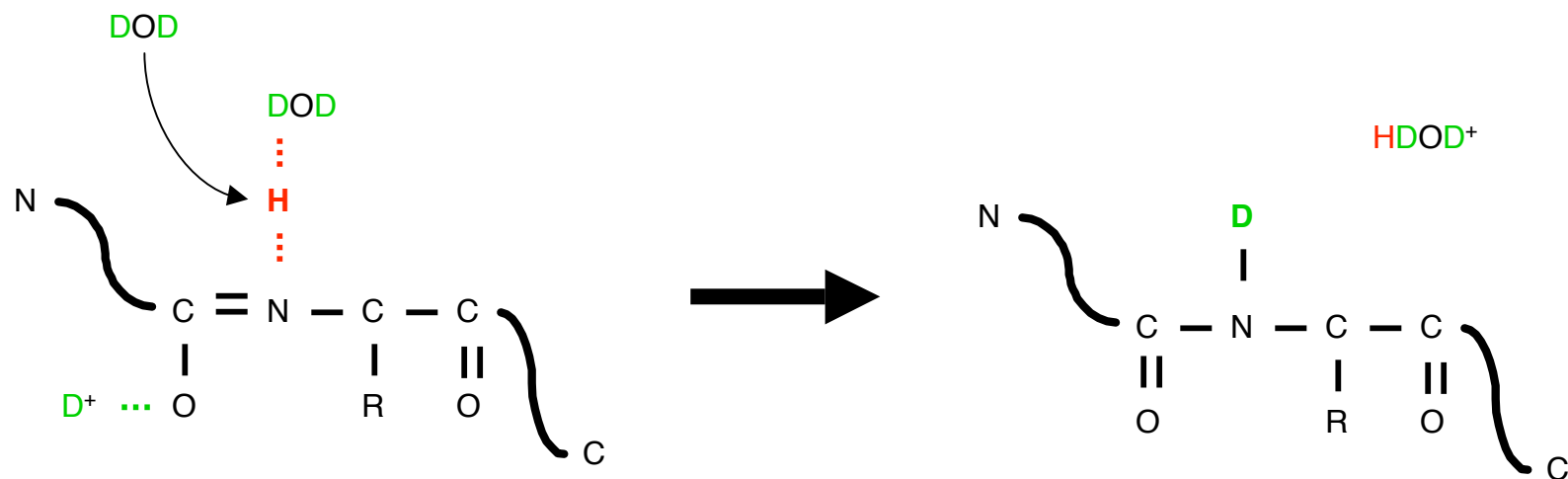
# Protein Unfolding



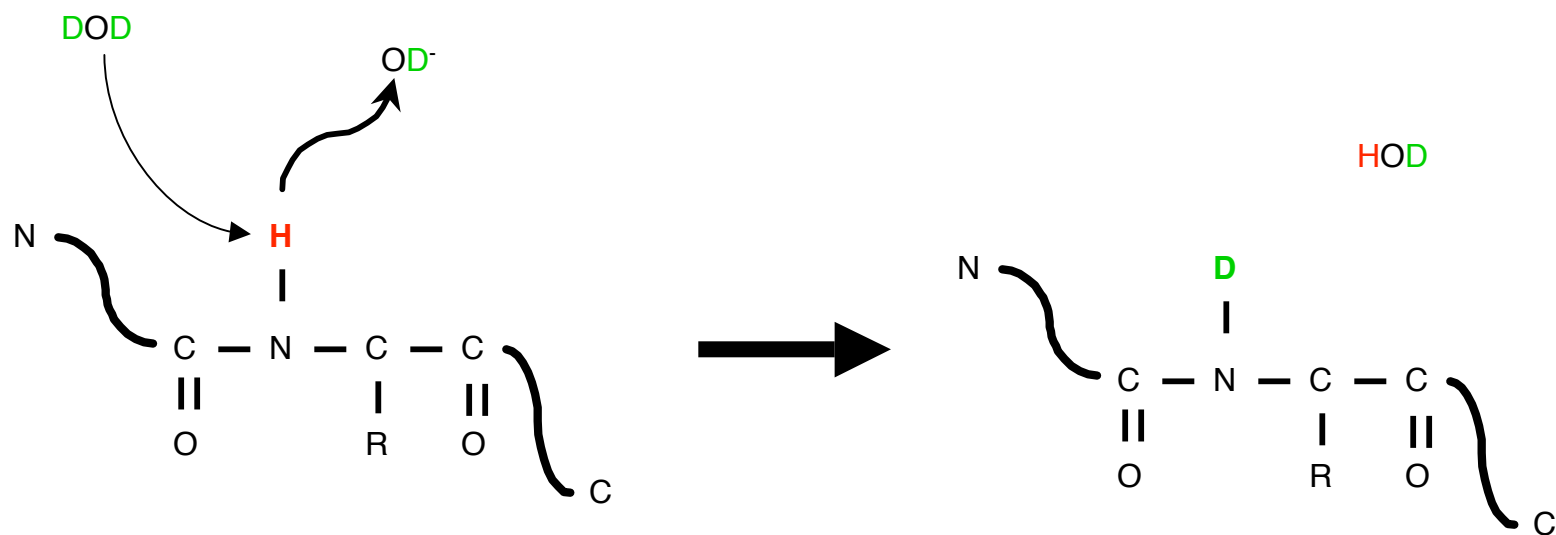
# Acid Catalysis



## Acid Catalysis

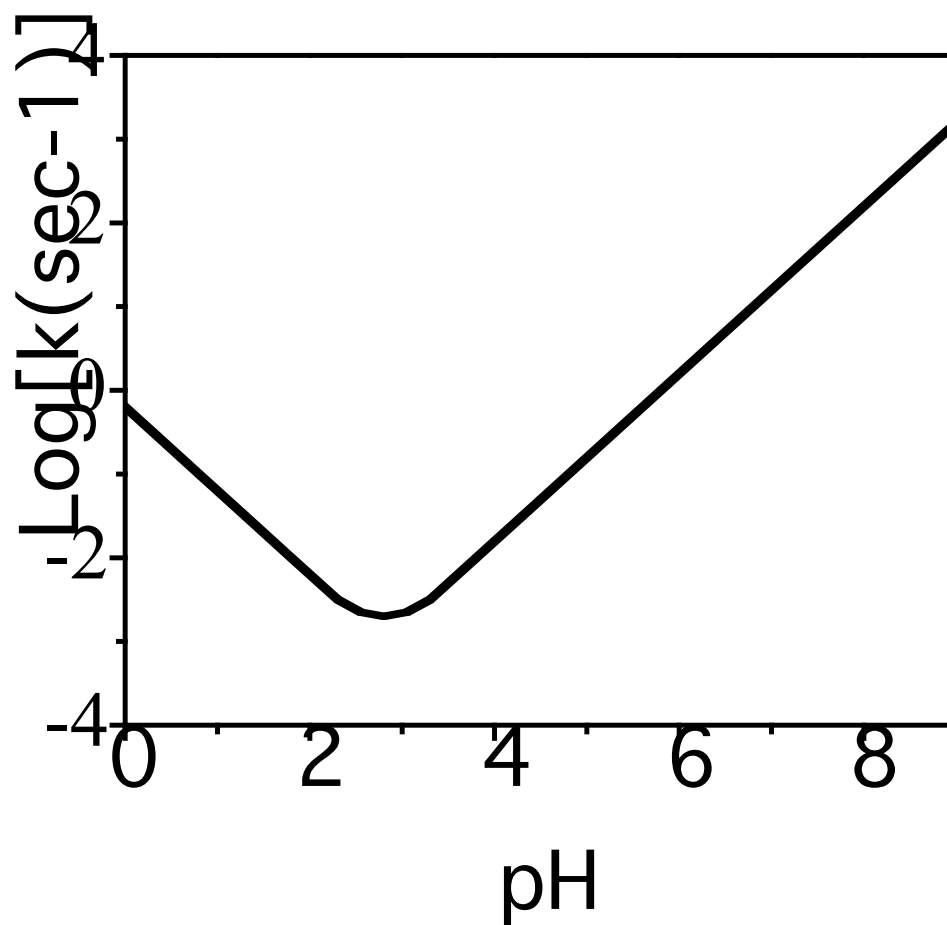


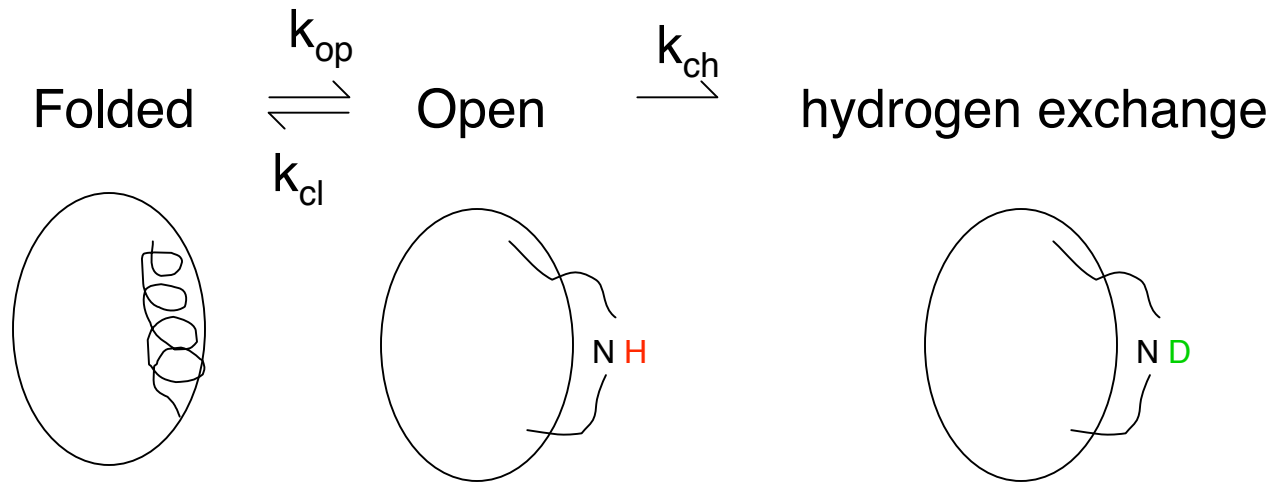
## Base Catalysis





Amide Proton Exchange is pH dependent





EX<sub>1</sub> mechanism:  $k_{ch} > k_{cl}$

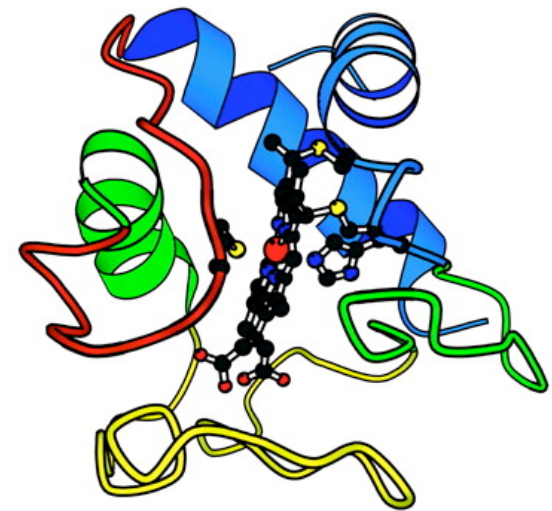
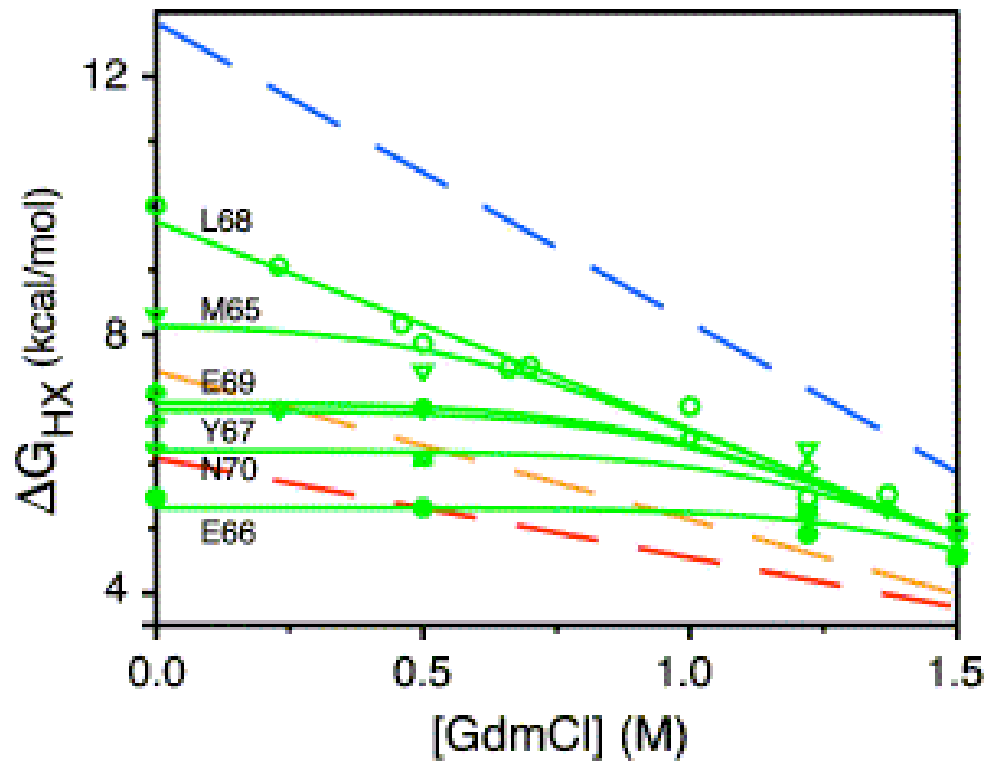
$$k_{ex} = k_{op}$$

EX<sub>2</sub> mechanism:  $k_{ch} < k_{cl}$

$$k_{ex} = K_{(op/cl)} \cdot k_{ch}$$

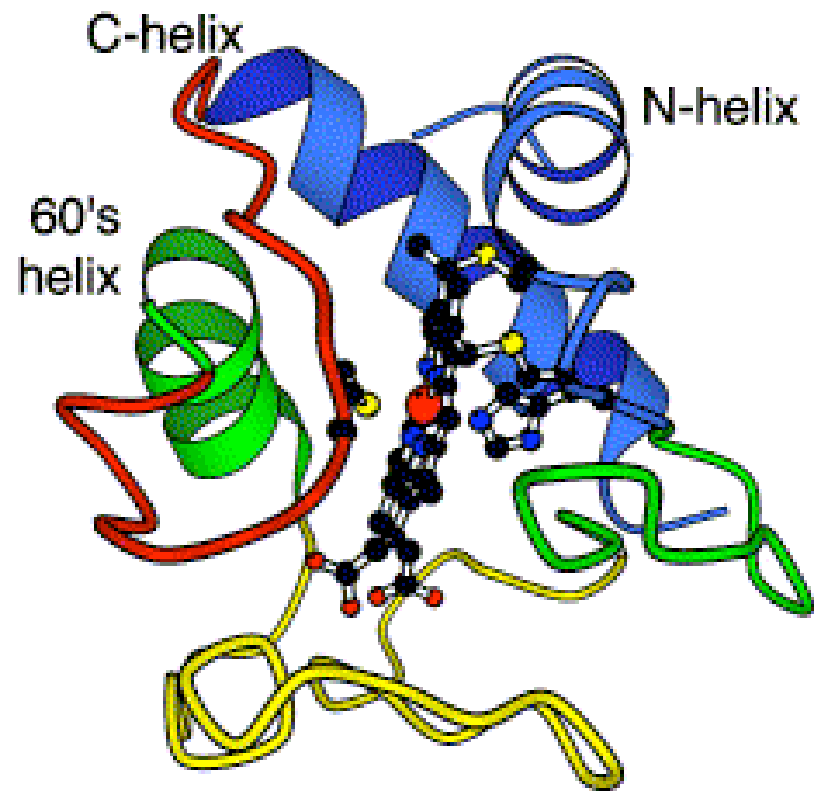
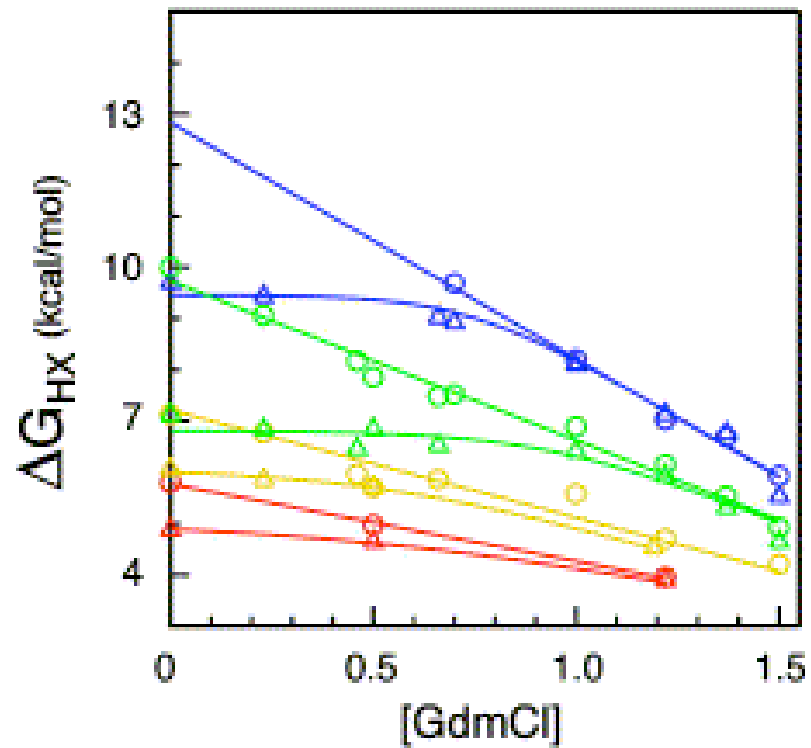
“pH dependent”

# Cooperative Unfolding



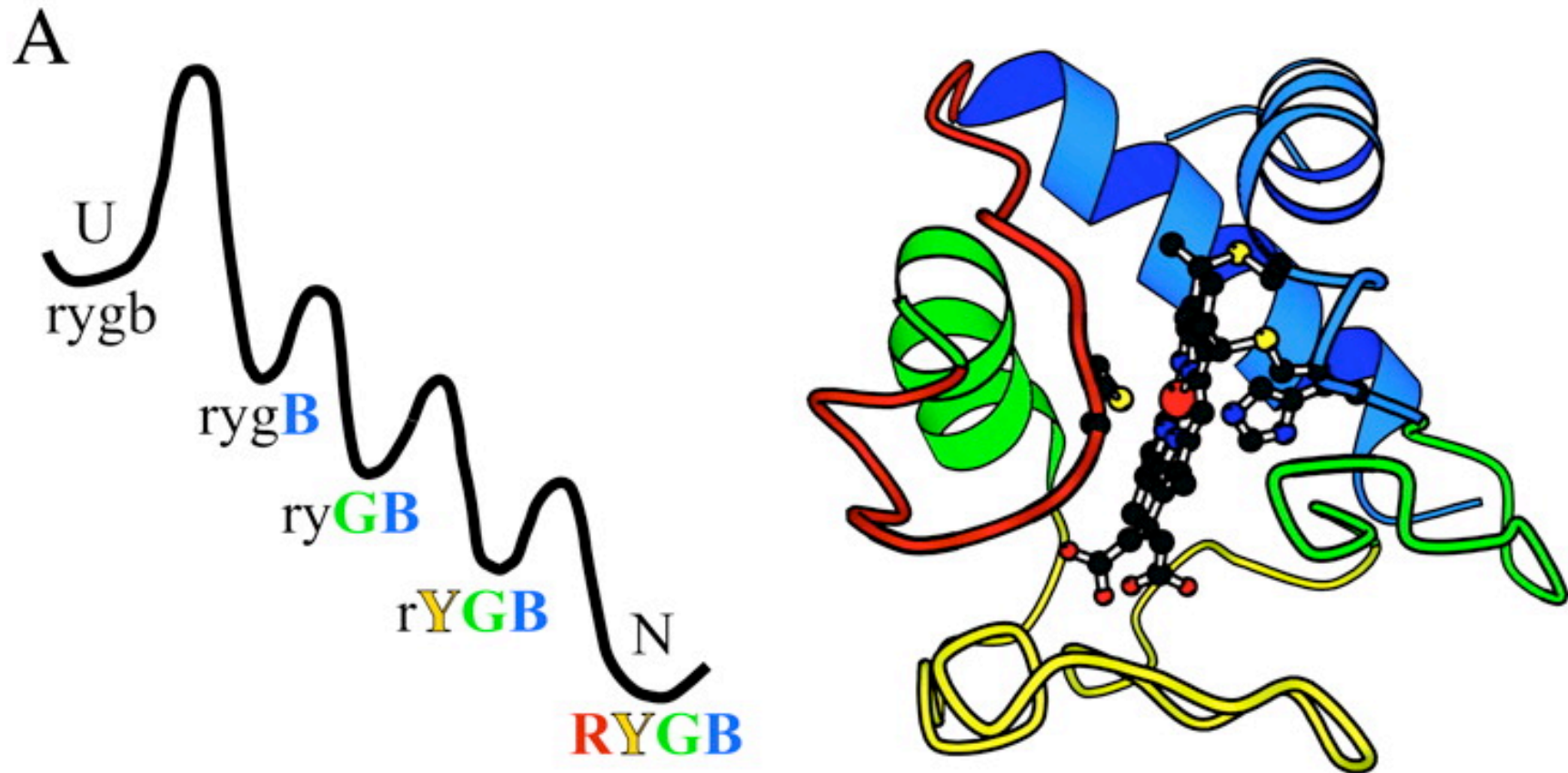
Cytochrome C

# Cooperative Unfolding

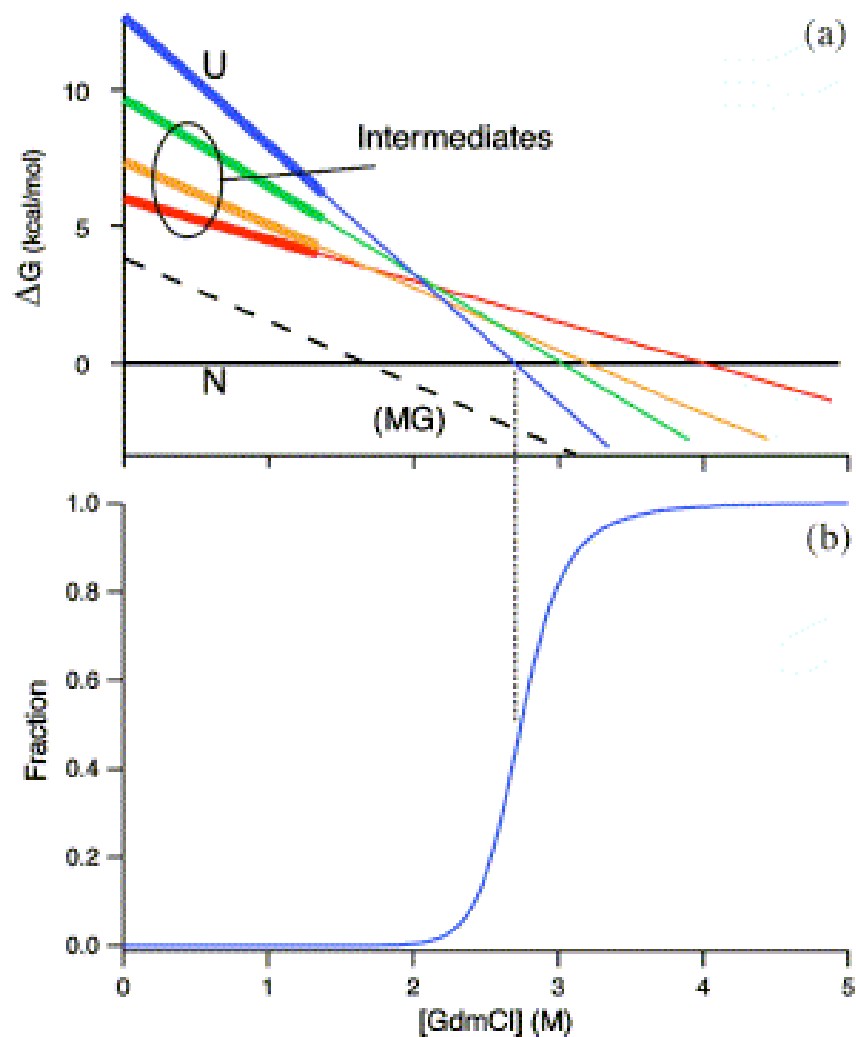


Cytochrome C

# Cytochrome C Folding Pathway



# Multi-state vs Two State Protein Unfolding



Englander, *et. al.* 2002

## NMR H/D exchange

Individual amide proton exchange rates

Sensitive to subtle protein dynamics

Used extensively to study protein folding

## Problems

Need pure sample

Need high concentrations

Only small proteins

# Mass Spec H/D exchange?

David Smith, (Zhang *et al* 1993)

Sample need not be pure

Low sample concentrations

Large proteins

Macromolecular complexes

## Problems

Digestion coverage

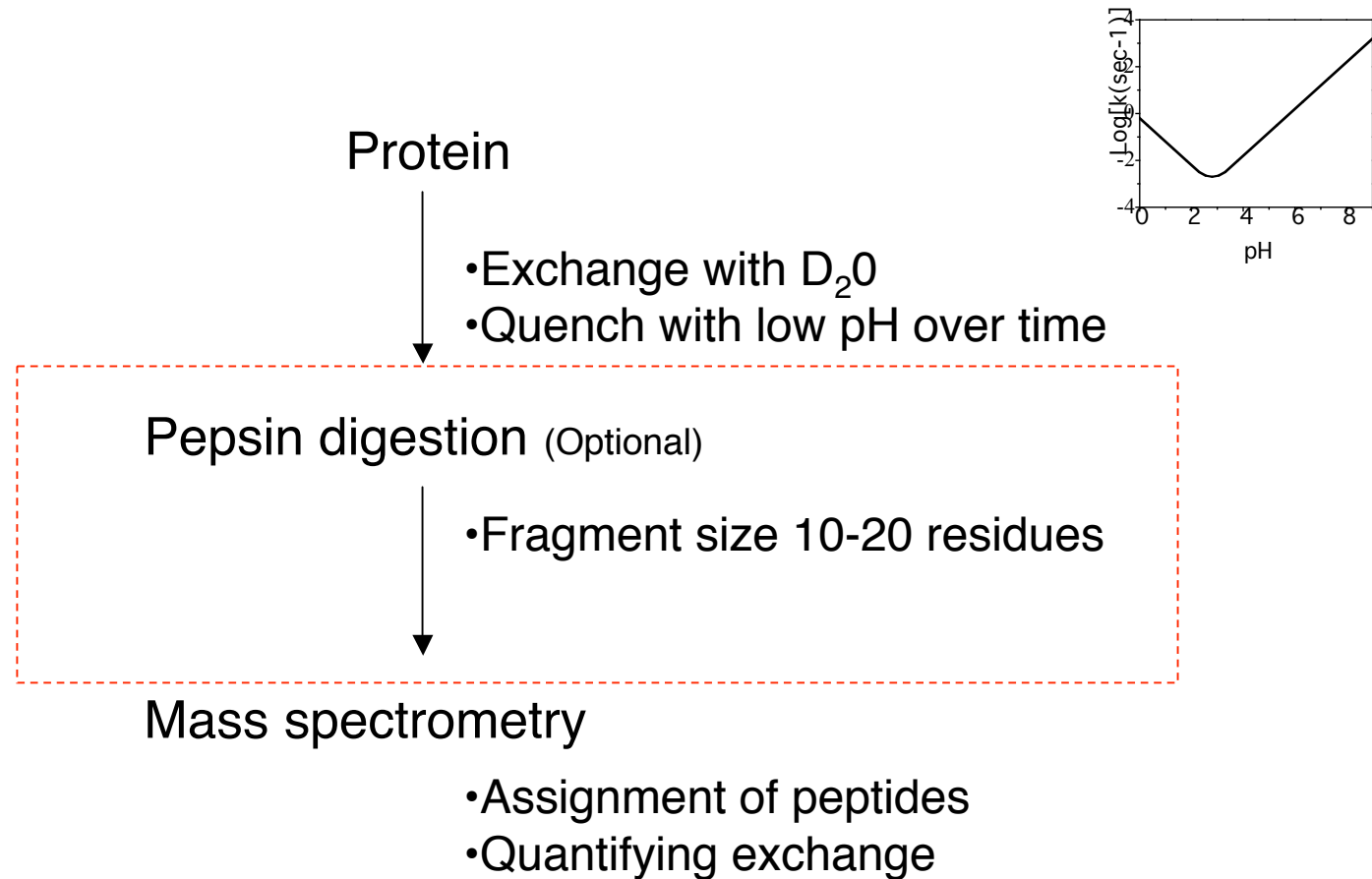
Exchange rate is averaged over the whole peptide

Buffer intolerance



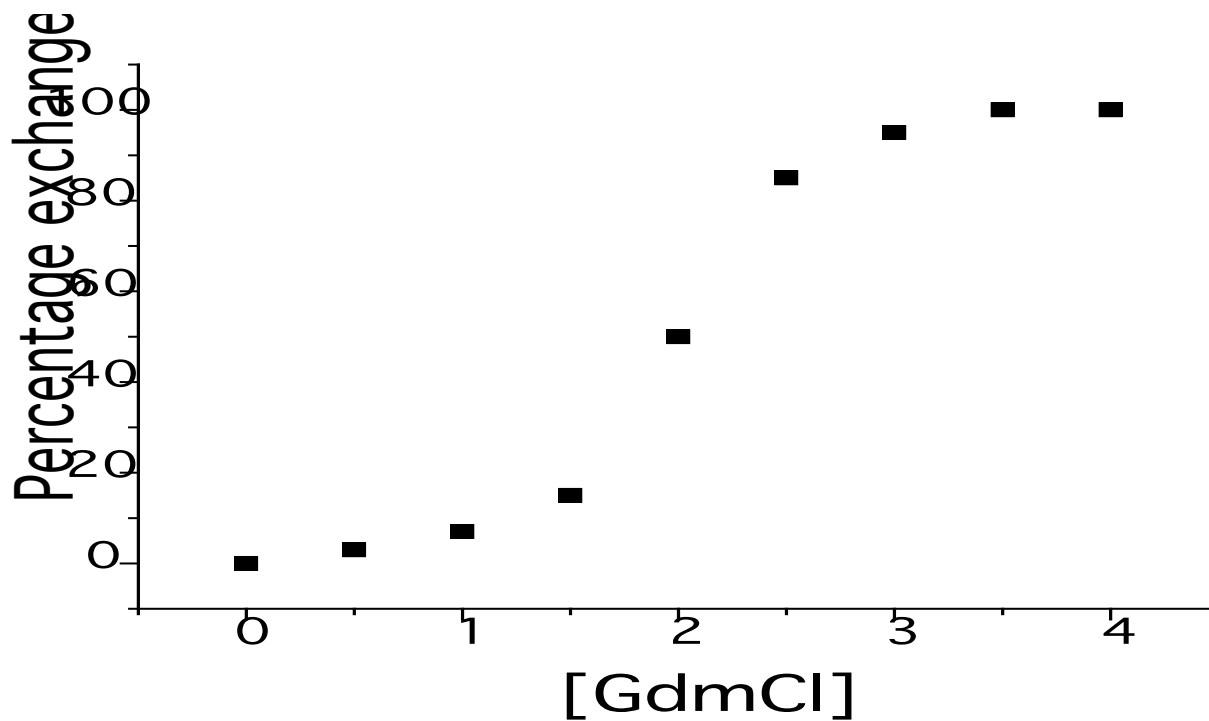
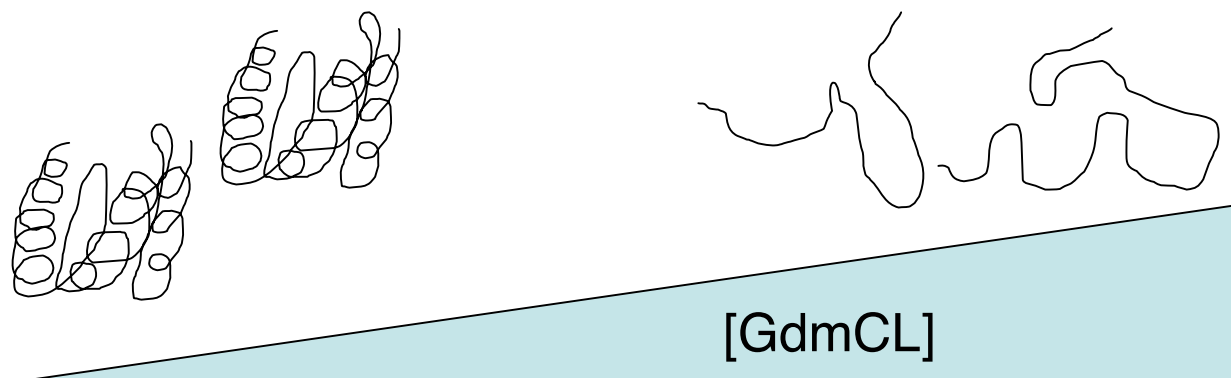
# H/D Exchange Experimental Protocol

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# Protein stability

“pulsed labeling H/D exchange”



# MALDI Analysis of Pulse labeled proteins “SUPREX”

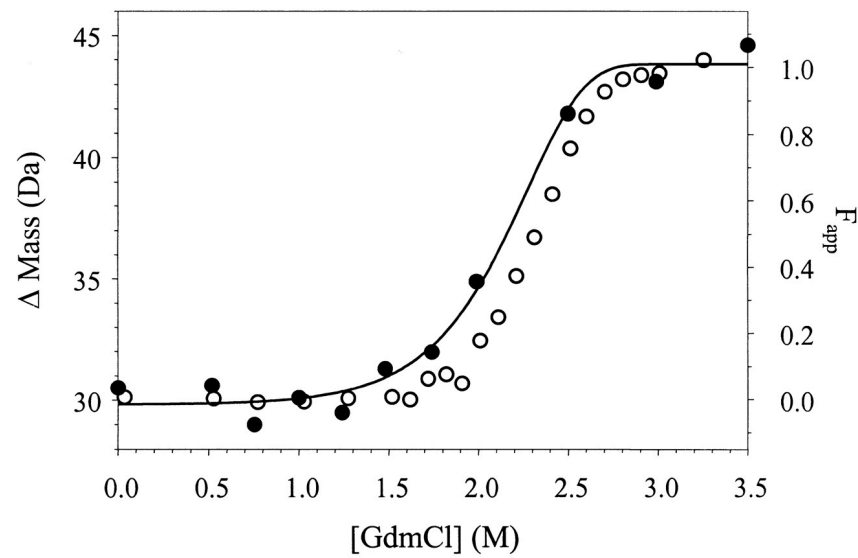
SUPREX; Stability of Unpurified Proteins from Rates of H/D Exchange

- Unpurified proteins
- Rapid analysis
- High throughput
- Protein stability in the cell

# SUPREX

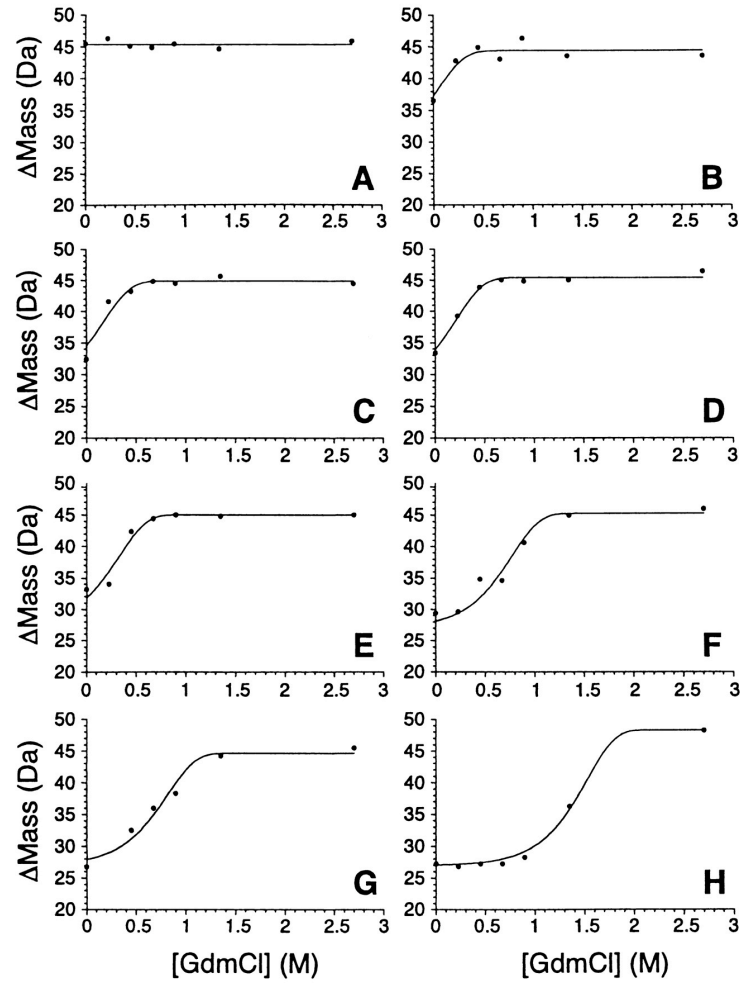
Suprex = ●

CD = ○



*4-oxalocrotonate tautomerase*

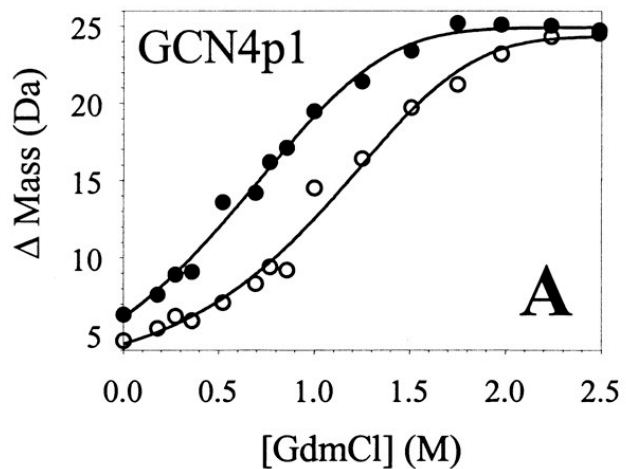
# SUPREX analysis of mutations and protein stability



Protein  
stability

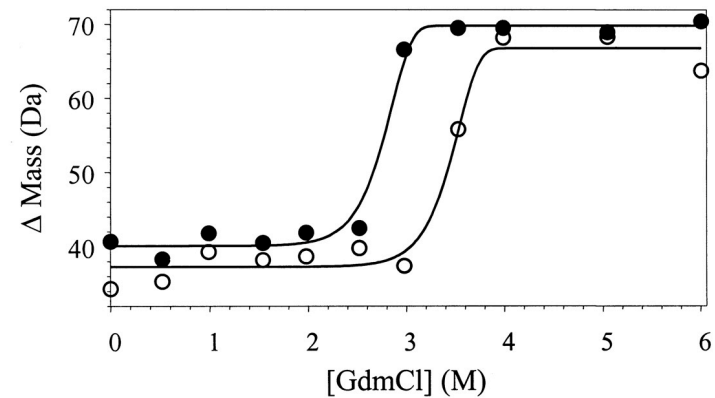
Maltose binding protein

# Association detected by SUPREX



10  $\mu$ M = ●

56  $\mu$ M = ○

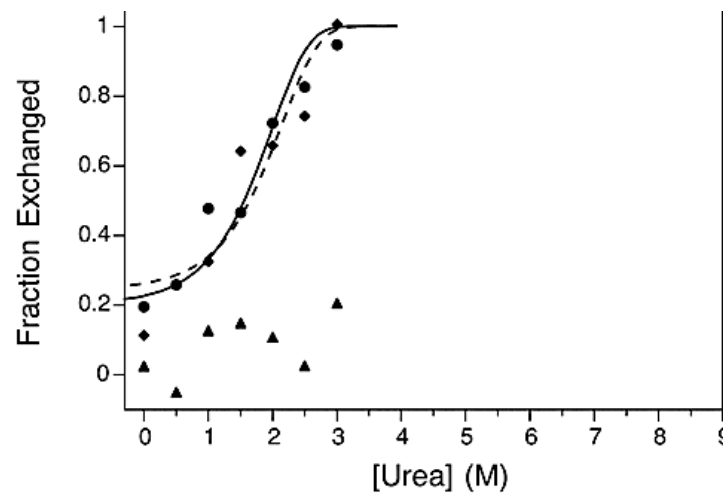
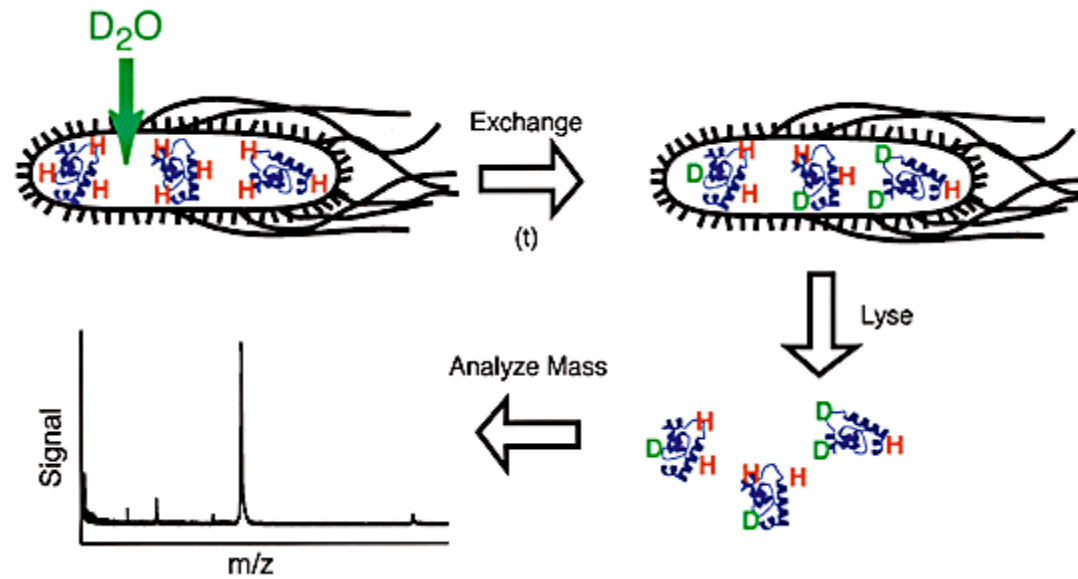


Trp repressor (TrpR)

- L-tryptophan = ●

+ L-tryptophan = ○

# Protein Stability in Cells



# MALDI Analysis of Pulse labeled proteins “SUPREX”

- Unpurified proteins
- Rapid analysis
- High throughput
- Protein stability in the cell

•?



*Continuous H/D Exchange*

*Protein folding*

*Protein interfaces*

*Quantitatively determine rates*

**?**

# H/D Exchange Experimental Protocol

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CA Tubes or Dimers



- Exchange with D<sub>2</sub>O
- Quench with low pH over time

Pepsin digestion

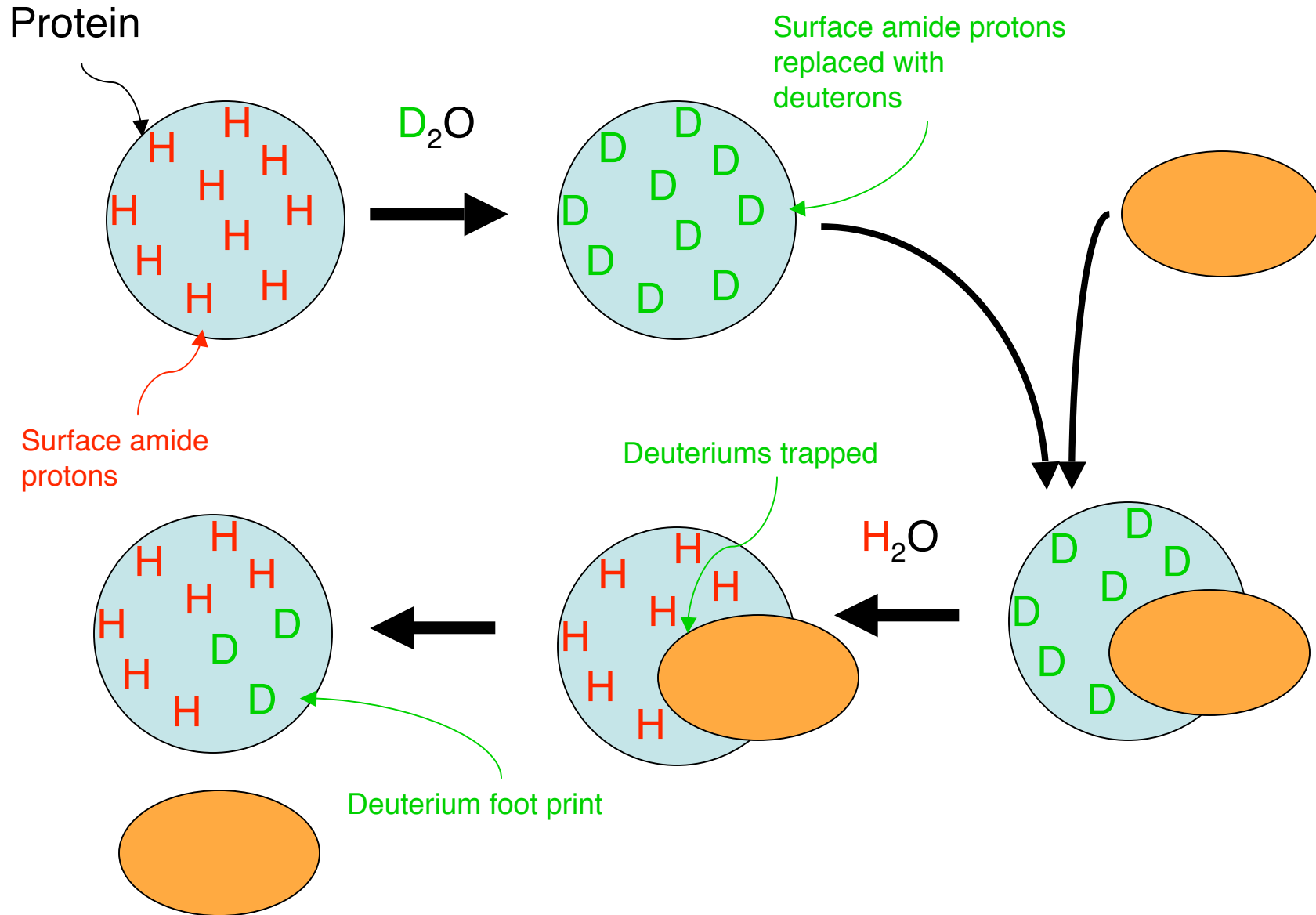


- Fragment size 10-20 residues

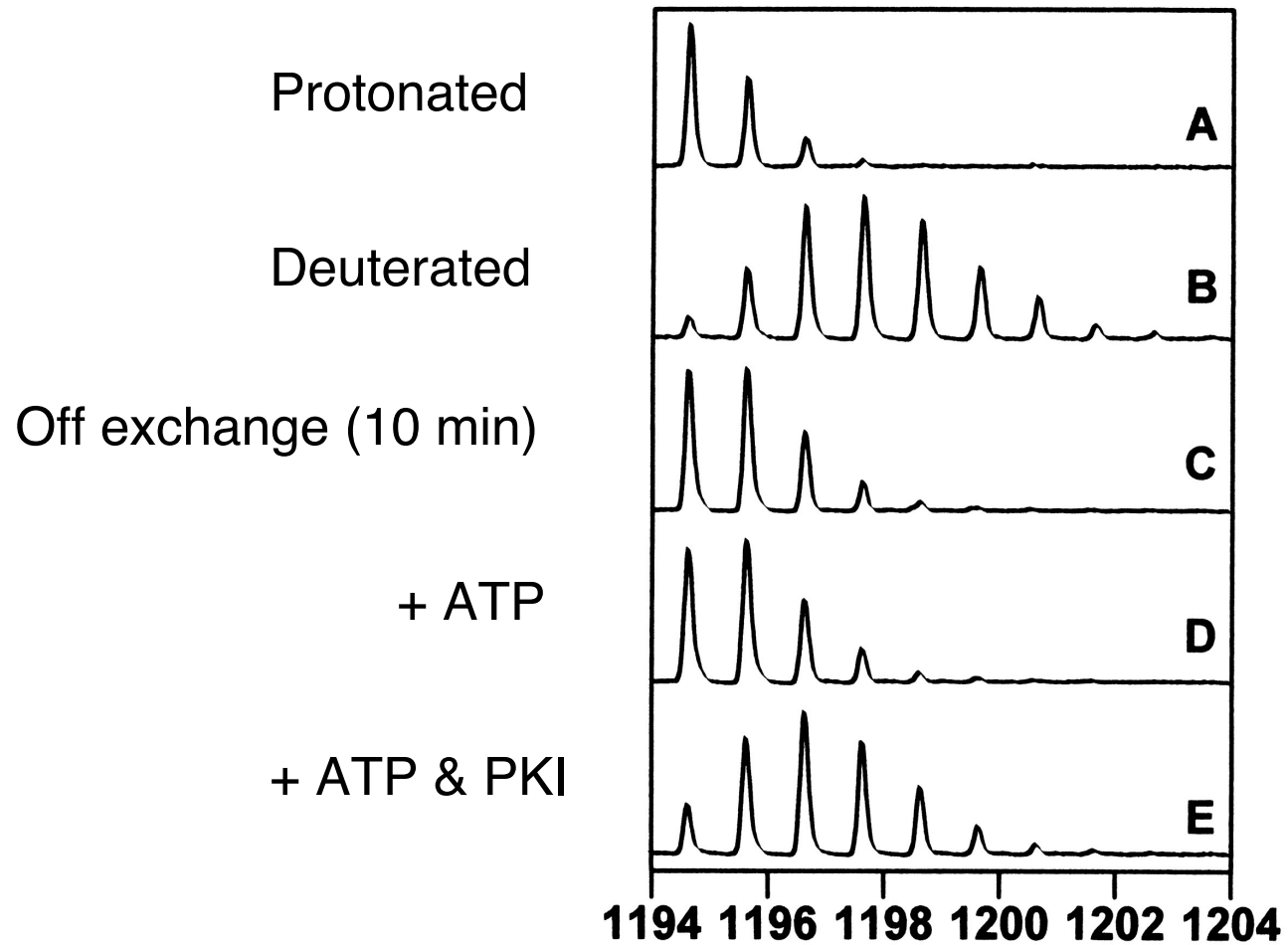
Mass spectrometry

- Assignment of peptides
- Quantifying exchange

# Identification of protein interaction interfaces

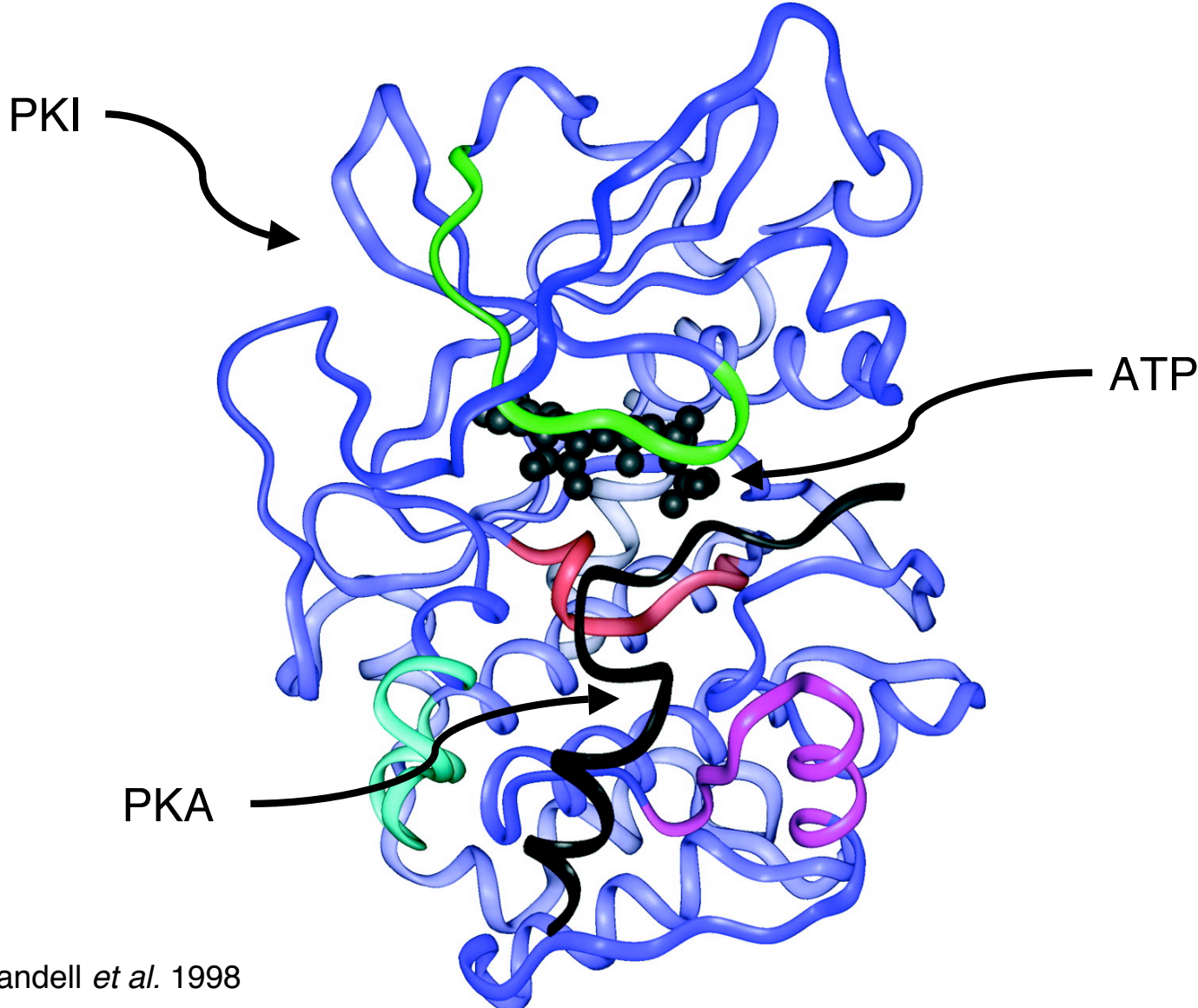


# Detection of PKI Interaction with PKA



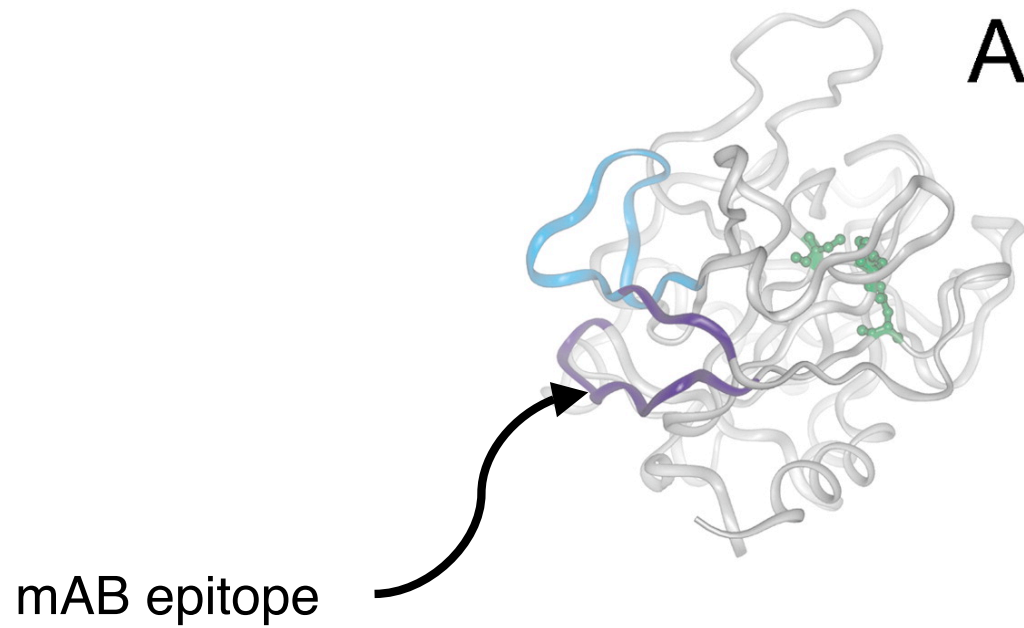
Mandell *et al.* 1998

# PKI, PKA, and ATP



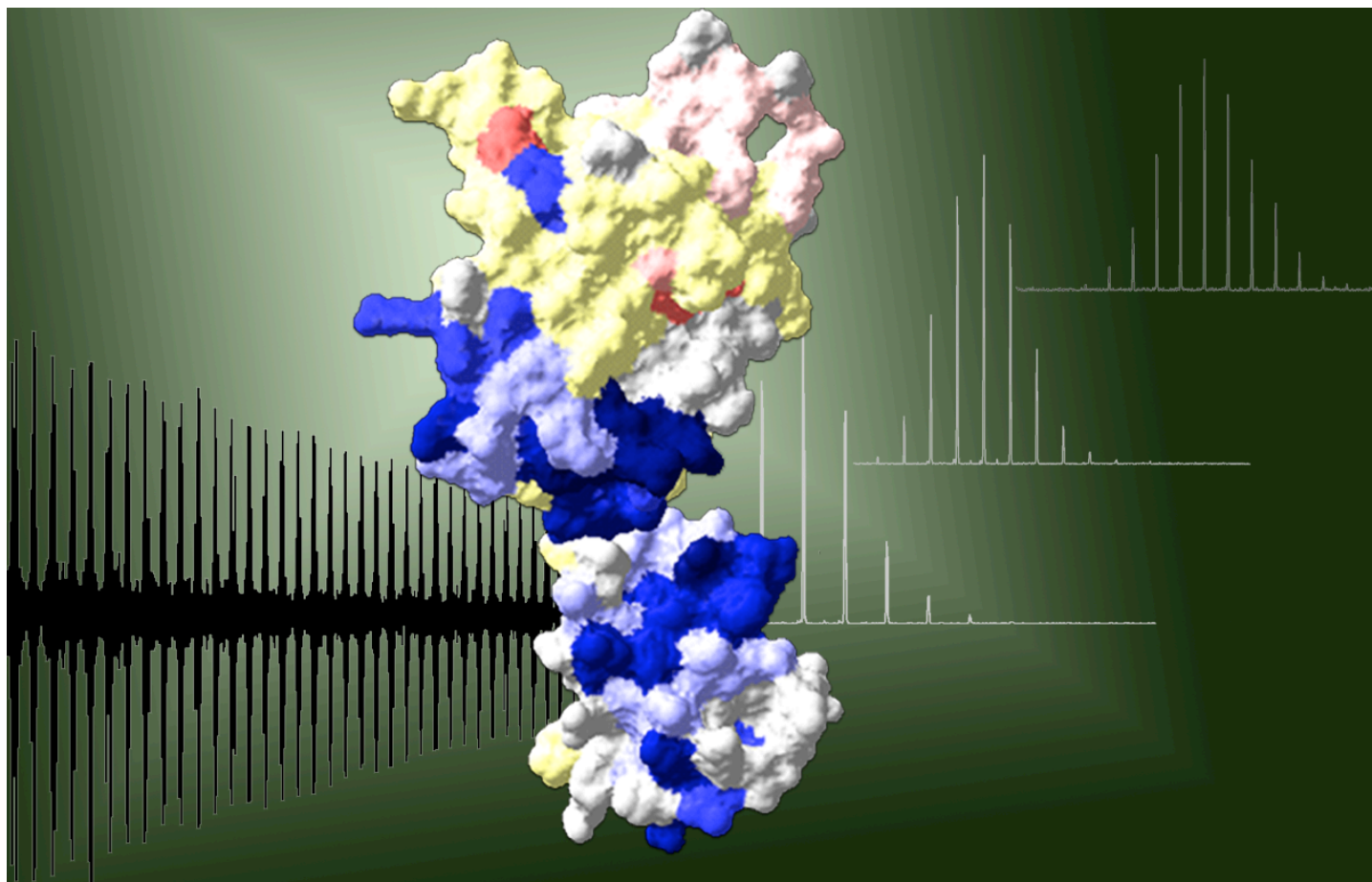
Mandell *et al.* 1998

# Epitope mapping of a monoclonal antibody against thrombin by H/D-exchange



# Novel Approaches for Understanding Virus Assembly and Dynamics.

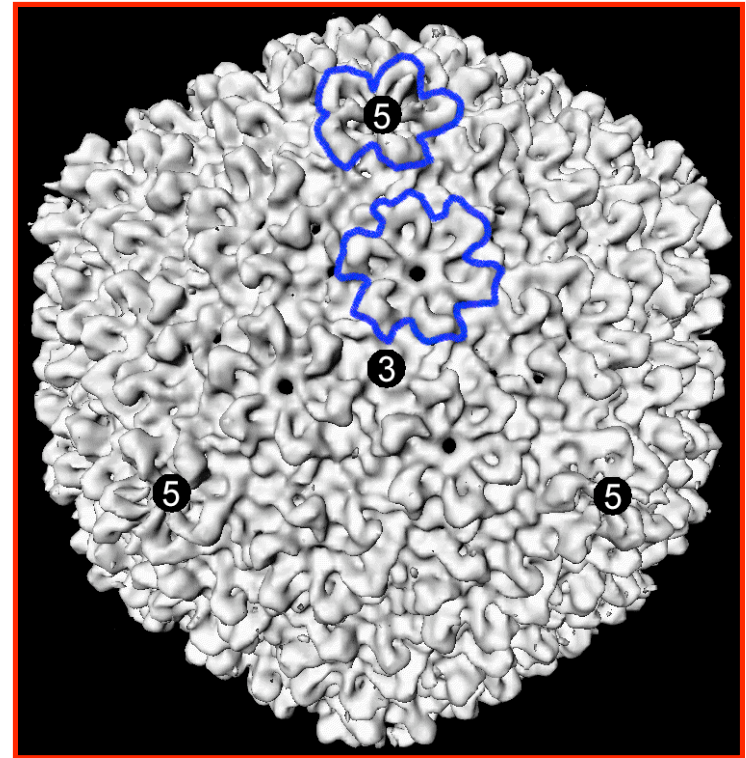
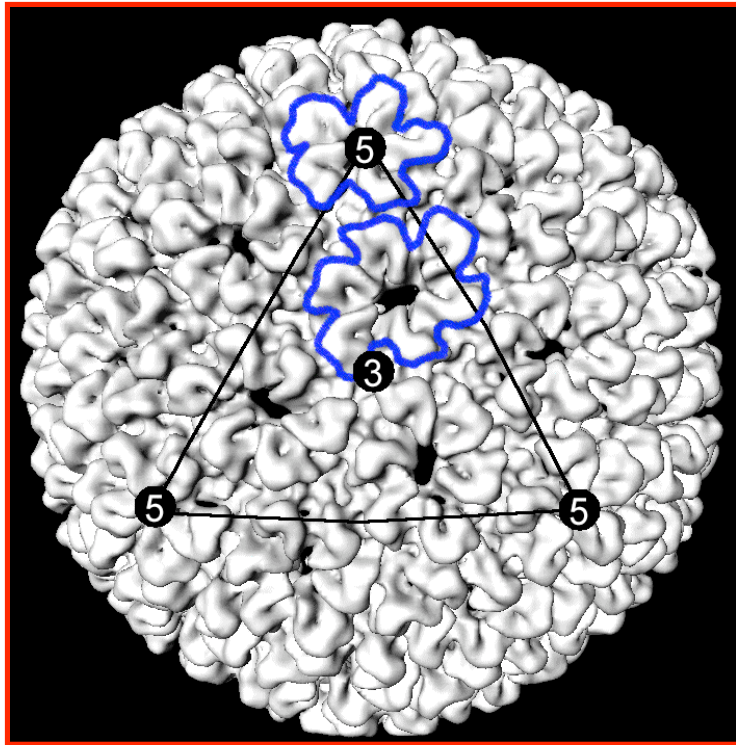
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Lanman *et al.* 2003

## Image Reconstructions of Procapsid and Mature Virion

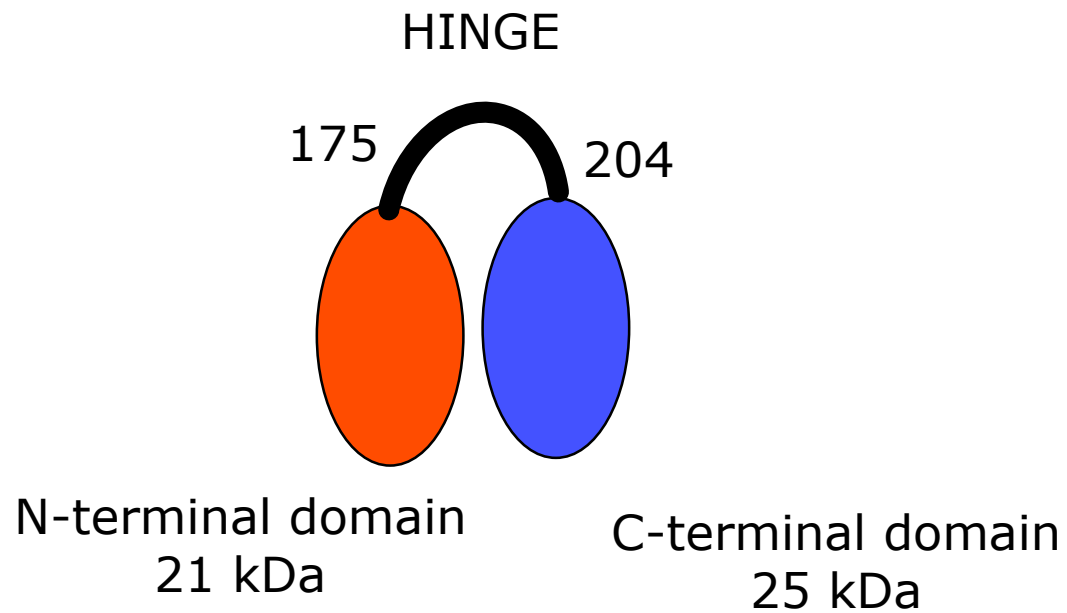
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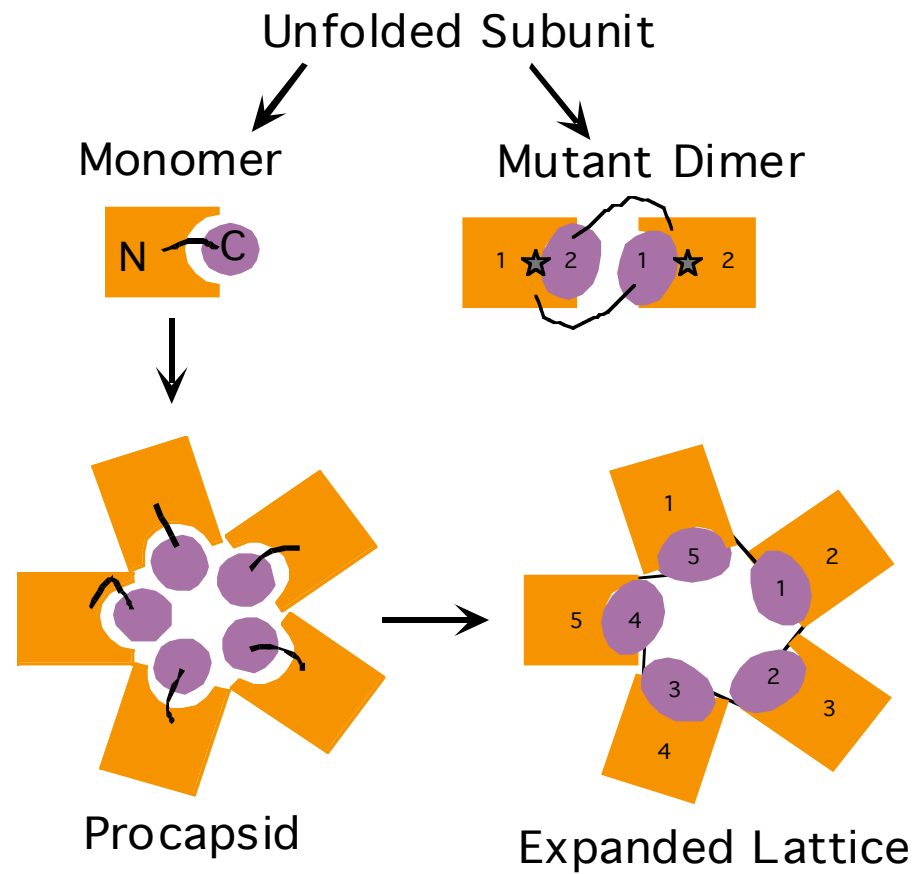
# The Coat Protein Subunit has a Two Domain Structure

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# A domain-shuffling model for capsid expansion

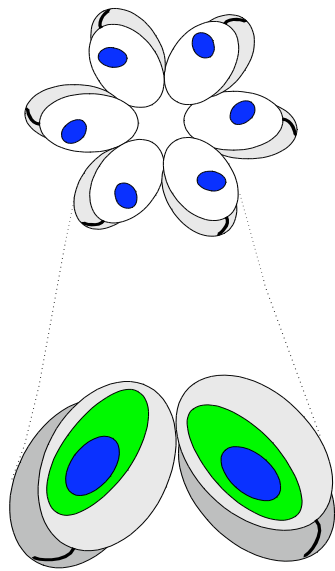
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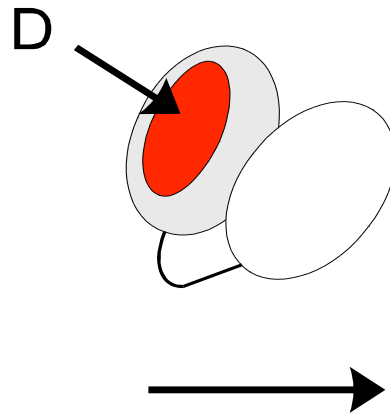
# The Model Predicts Trapping of Deuterium during Expansion

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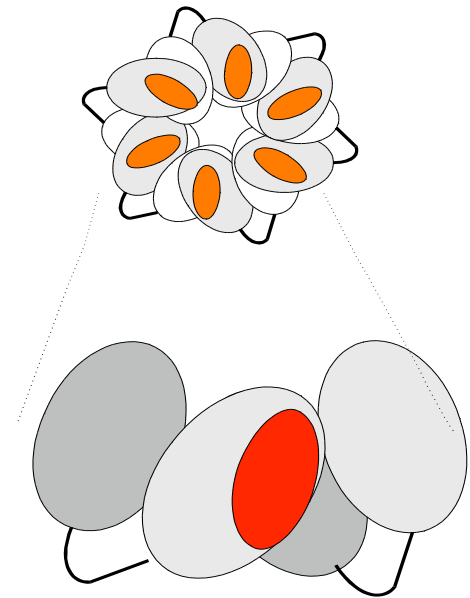
Procapsid



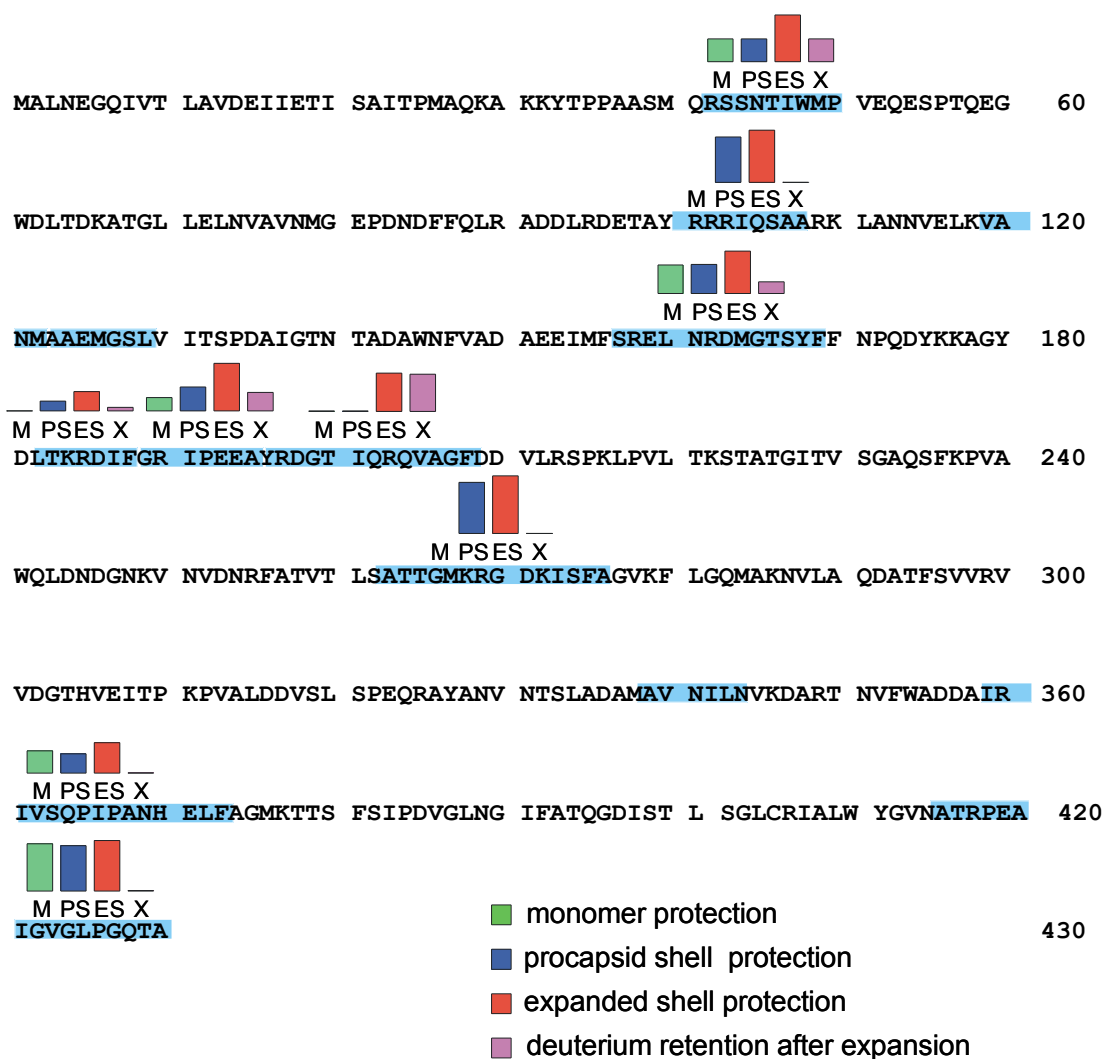
Transition State



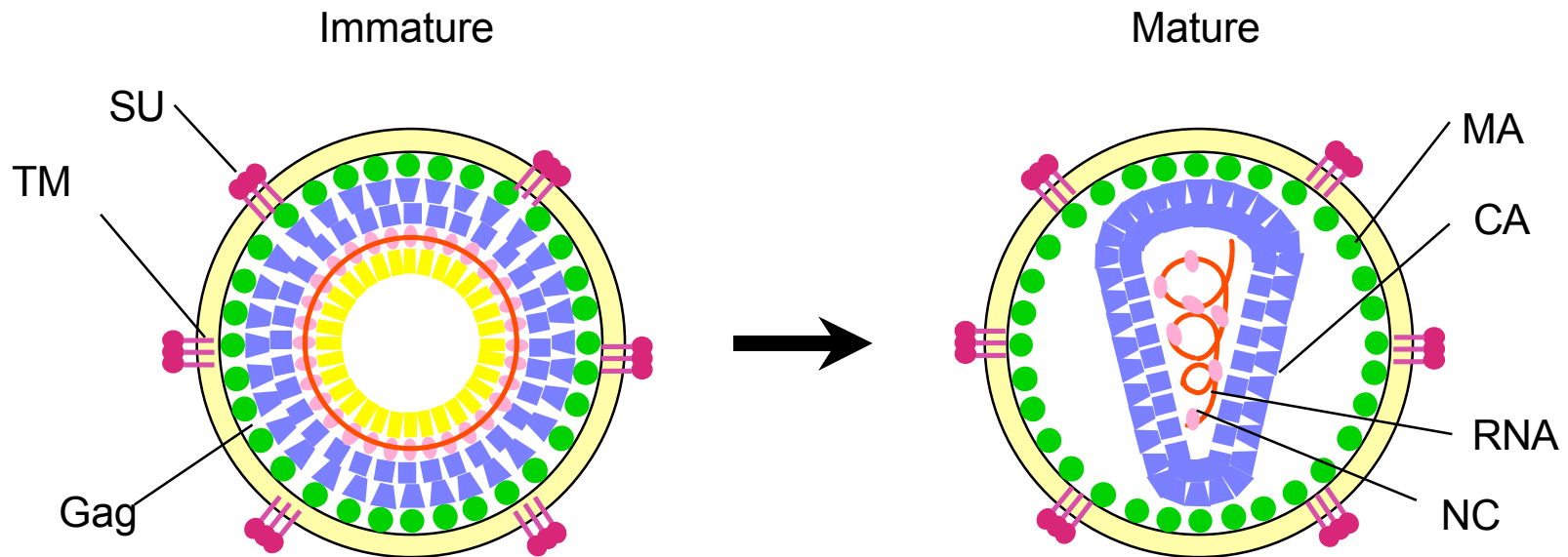
Expanded



# Changes in Exchange Protection during Assembly & Maturation



# HIV Assembly & Maturation

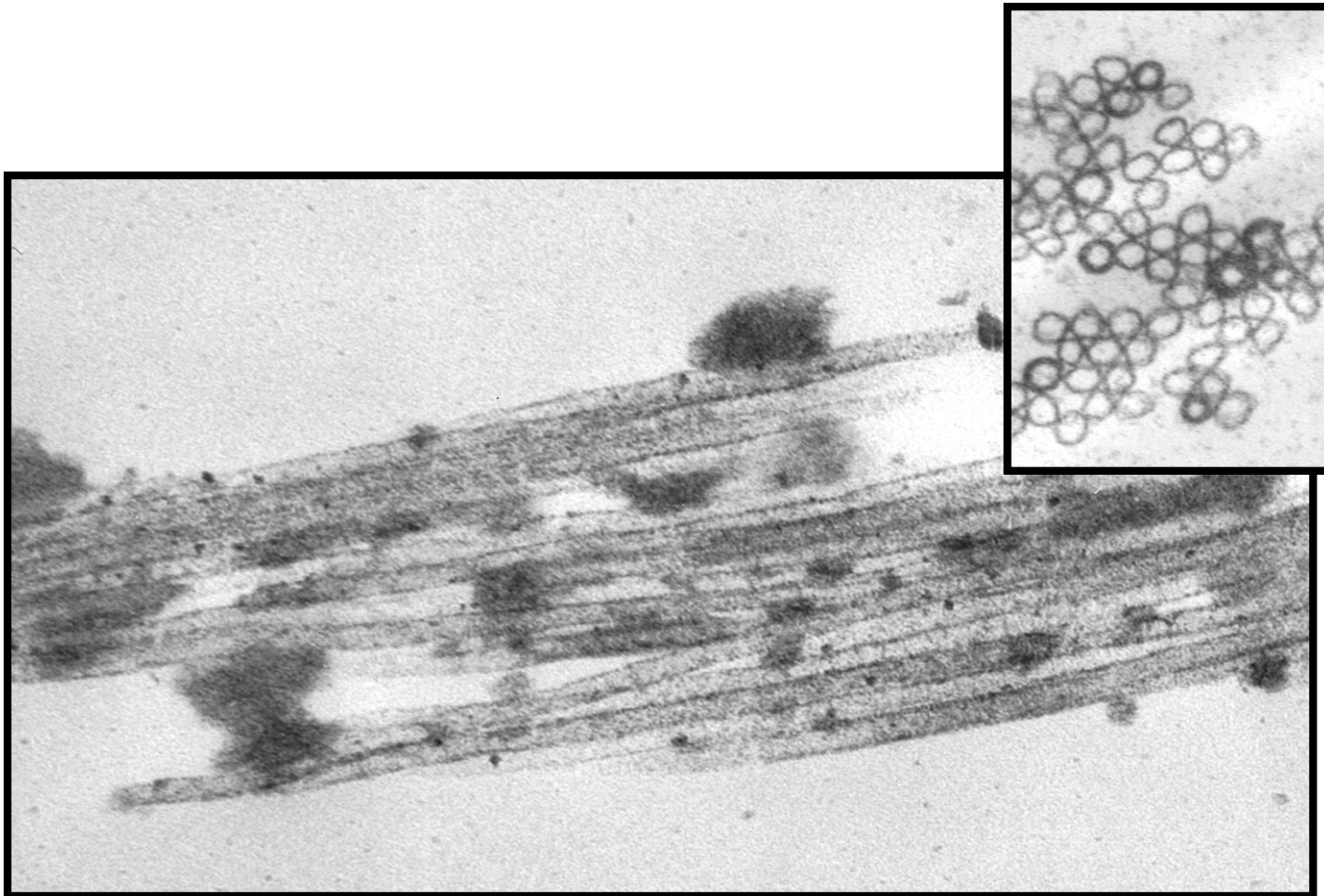


Gag



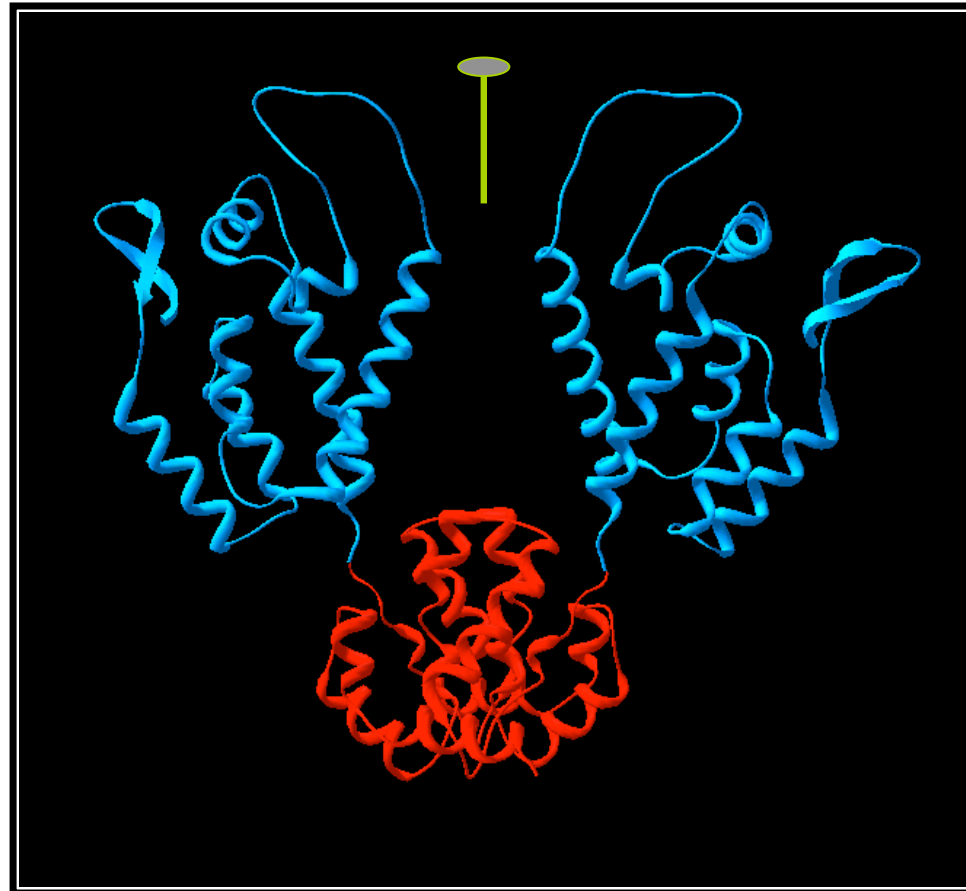
# Thin-Section EM Analysis of Assembly Products

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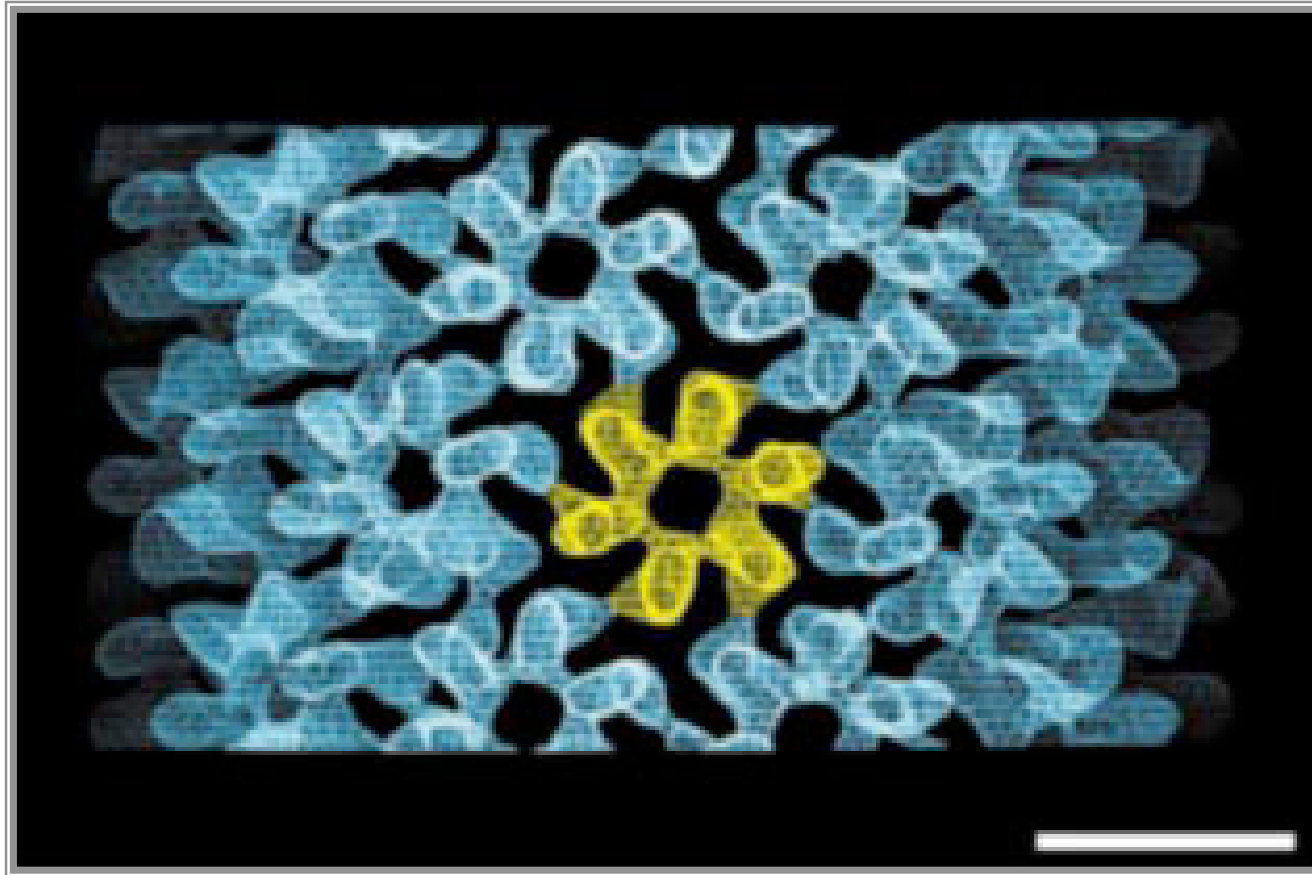
# CA is Comprised of Distinct N- and C-Terminal Structural Domains

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## CA Cylinders are Based on a Hexamer Lattice

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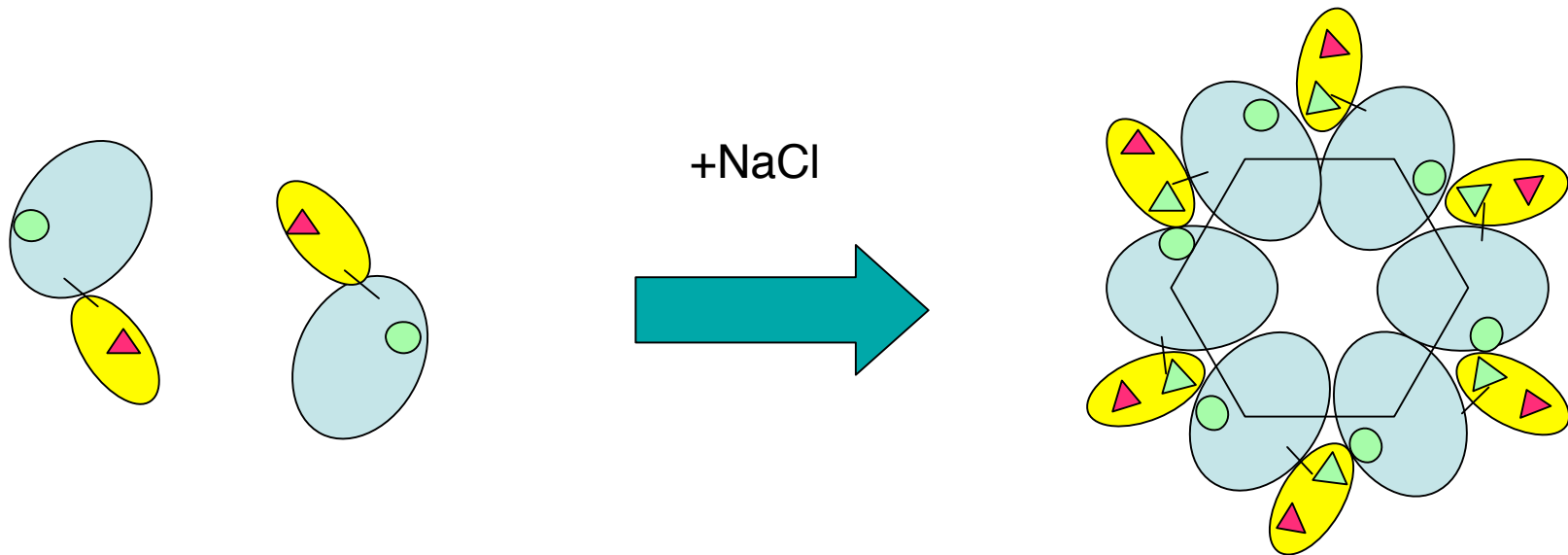


From Li et al, Nature 407:409 (2000)



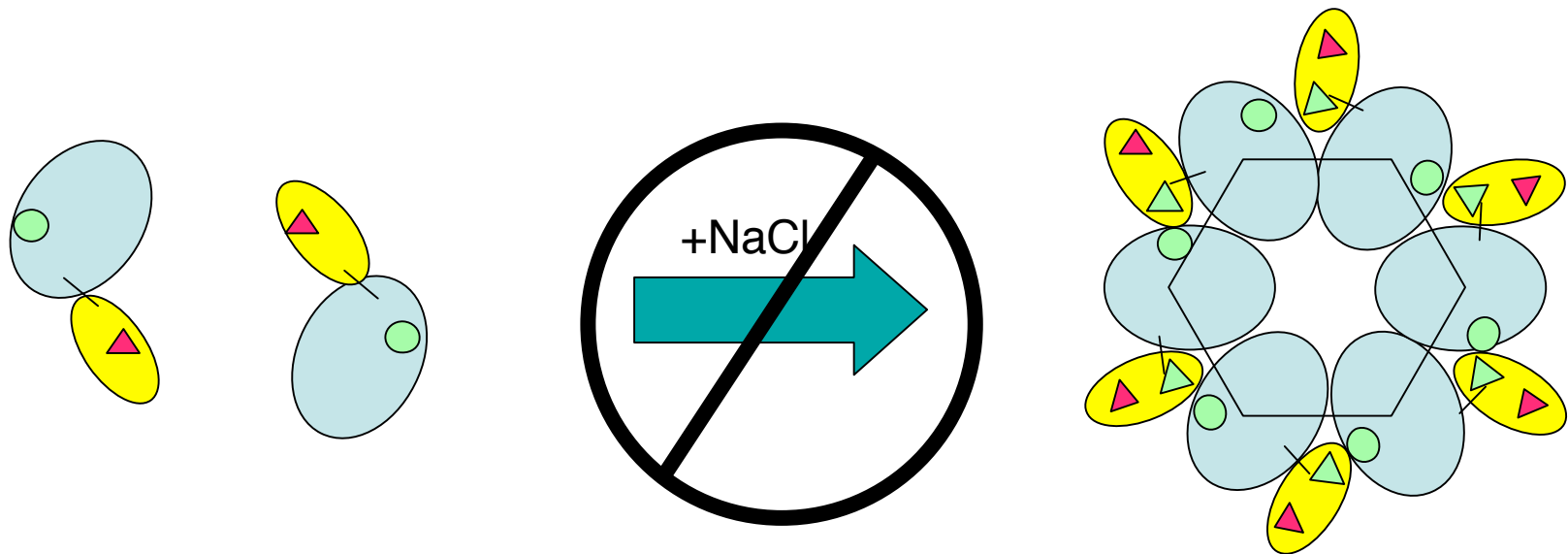
Hypothesis: The CA that does not form dimers, because of a mutation at the dimer interface should form hexamers at high NaCl concentrations

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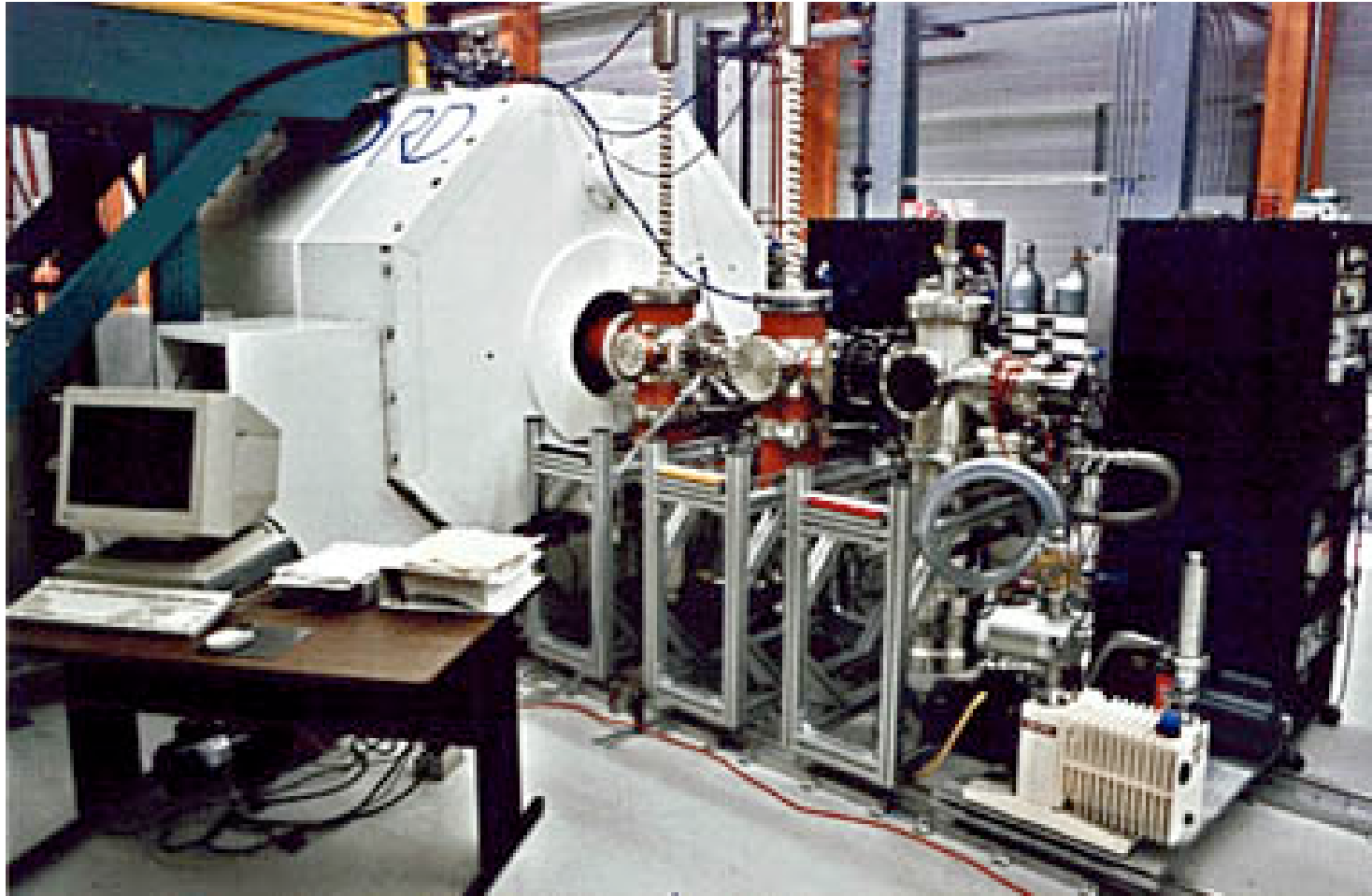
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CA does not form hexamers without C-domain (dimer) interaction.

## H/D exchange analyzed with FT-MS

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# H/D Exchange Experimental Protocol

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CA Tubes or Dimers



- Exchange with D<sub>2</sub>O
- Quench with low pH over time

Pepsin digestion

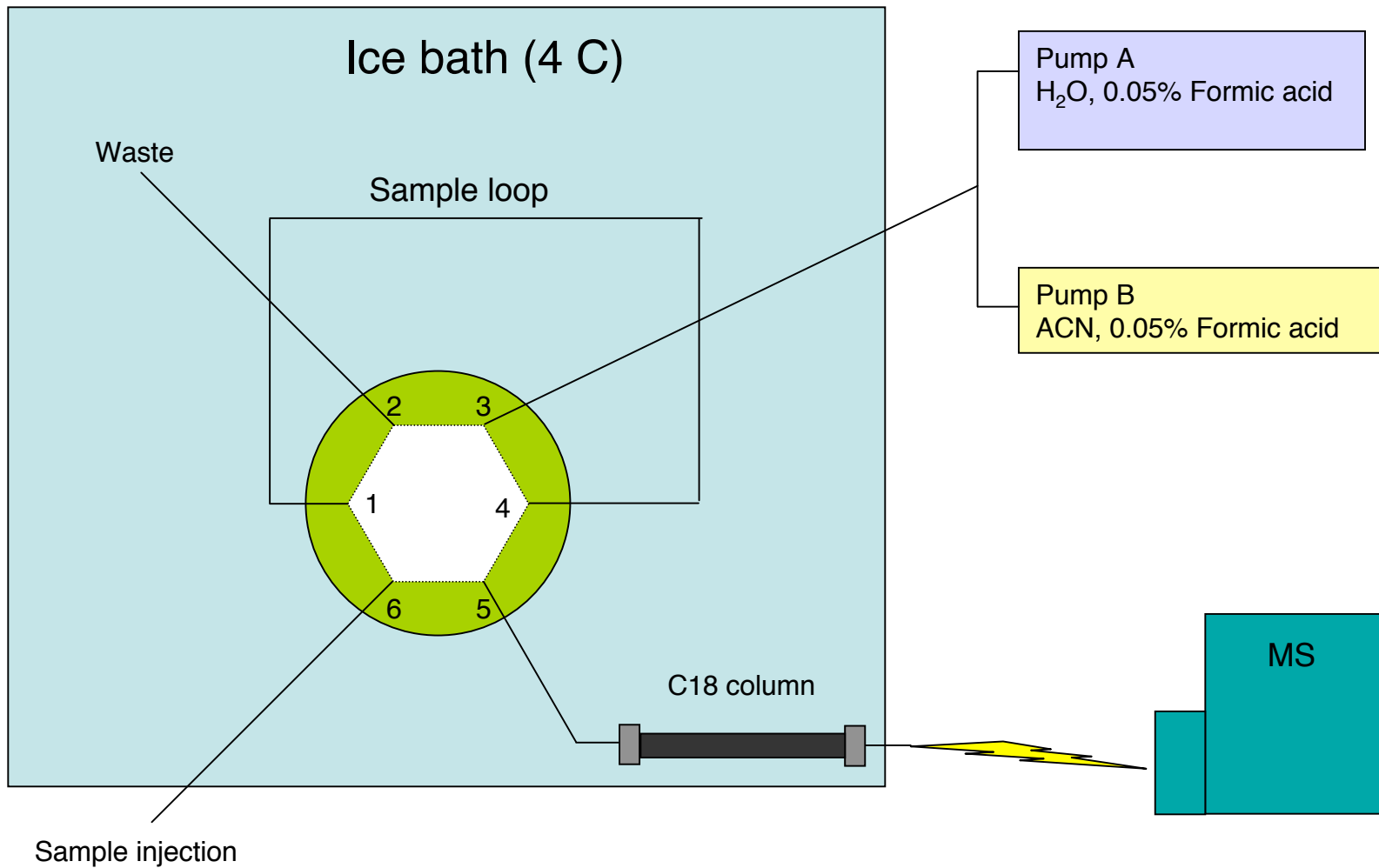


- Fragment size 10-20 residues

Mass spectrometry

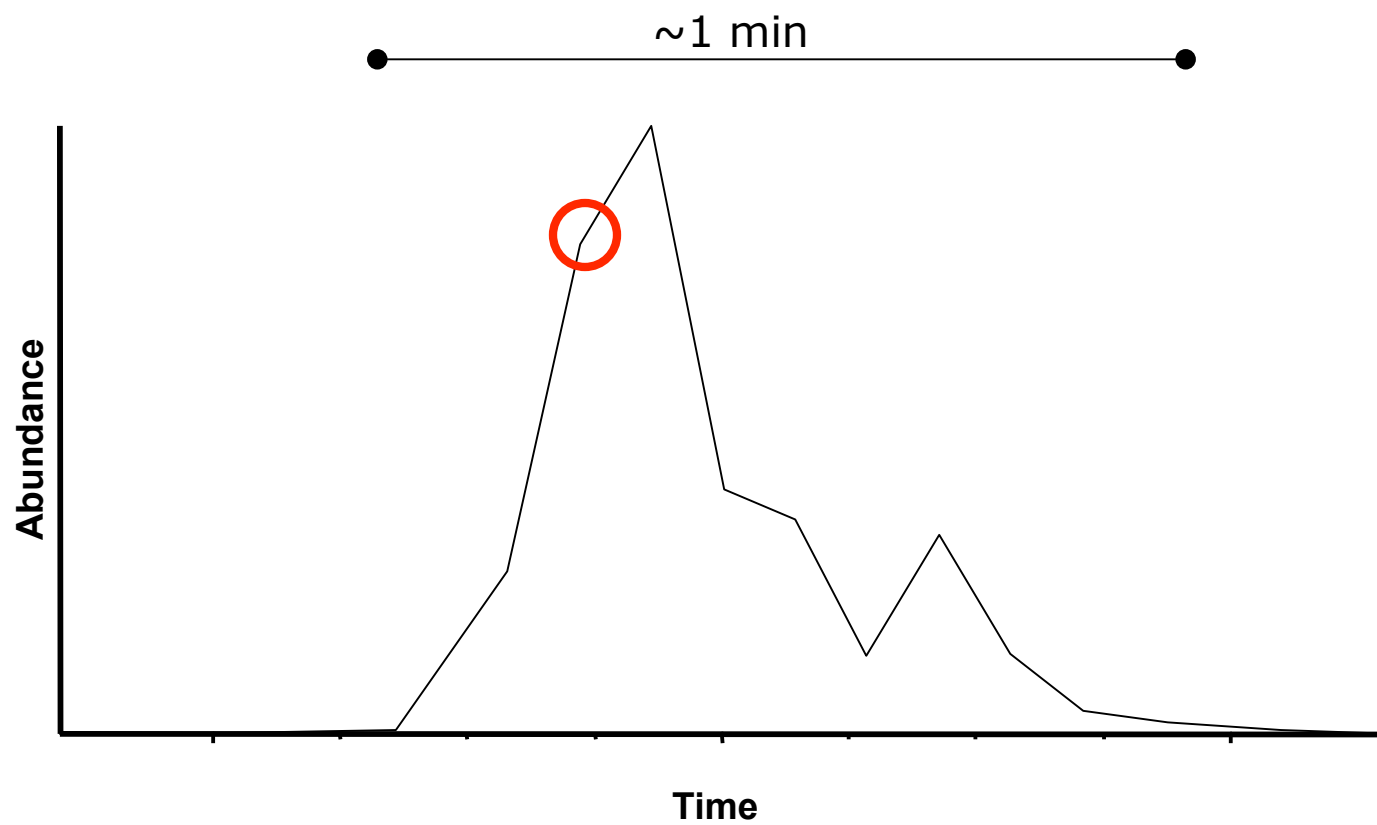
- Assignment of peptides
- Quantifying exchange

# ESI-MS Analysis of Peptides



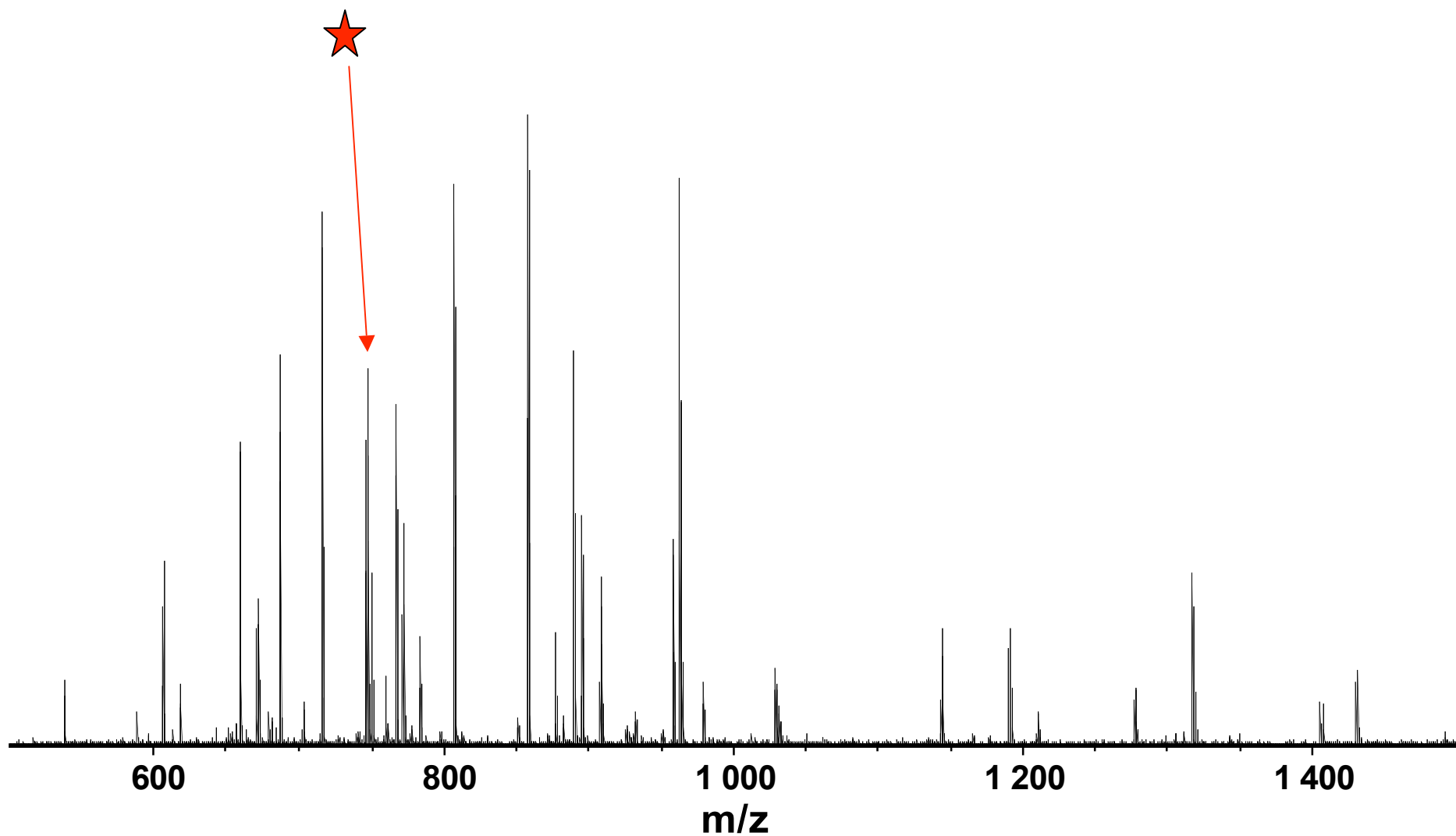
# Elution of Digest from C4 Column

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# Representative FT-MS Scan

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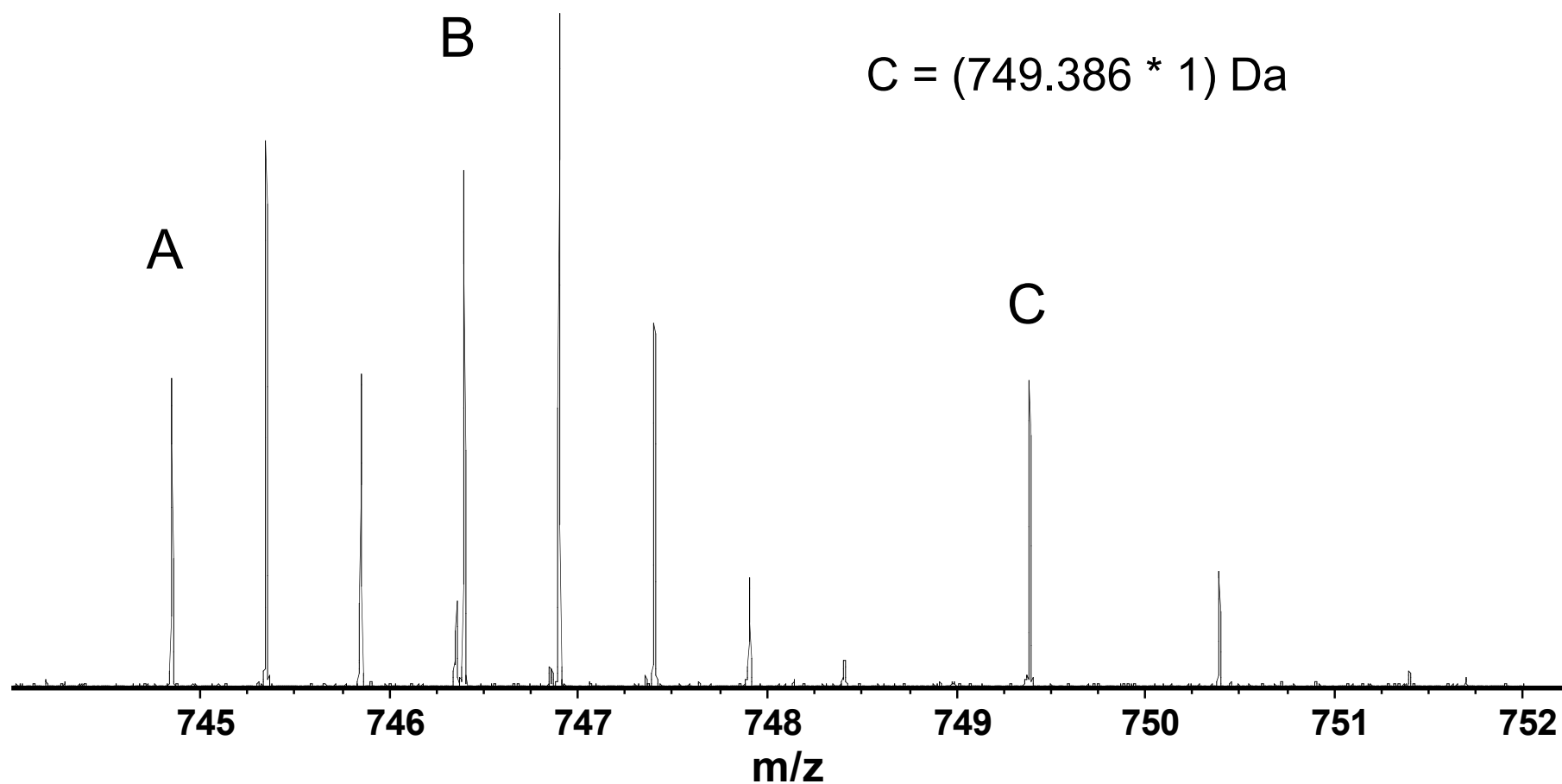
## Expanded View of 744-752 m/z Region

---

$$A = (744.842 * 2) \text{ Da}$$

$$B = (746.390 * 2) \text{ Da}$$

$$C = (749.386 * 1) \text{ Da}$$





# With High Resolution Peptides of Similar m/z can be Analyzed

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744.842 within 1 ppm  
746.390 within 0.5 ppm



## Peptides Covering 95% of the Sequence have Been Assigned

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1

PIVQNLQGQM VHQAISPRTL NAWVKVVEEK AFSPEVIPMF SALSEGATPQ DLNTMLNTVG

□

61

GHQAAMQMLK ETINEEAAEW DRLHPVHAGP IAPGQMREPR GSDIAGTTST LQEIQIGWMTH

121

NPPIPVGGEIY KRWIILGLNK IVRMYSPTSI LDIRQGPKEP FRDYVDRFYK TLRAEQASQE

□

□

181

VKNWMTETLL VQANANPDCKT ILKALGPGAT LEEMMTACQG VGGPGHKARV L

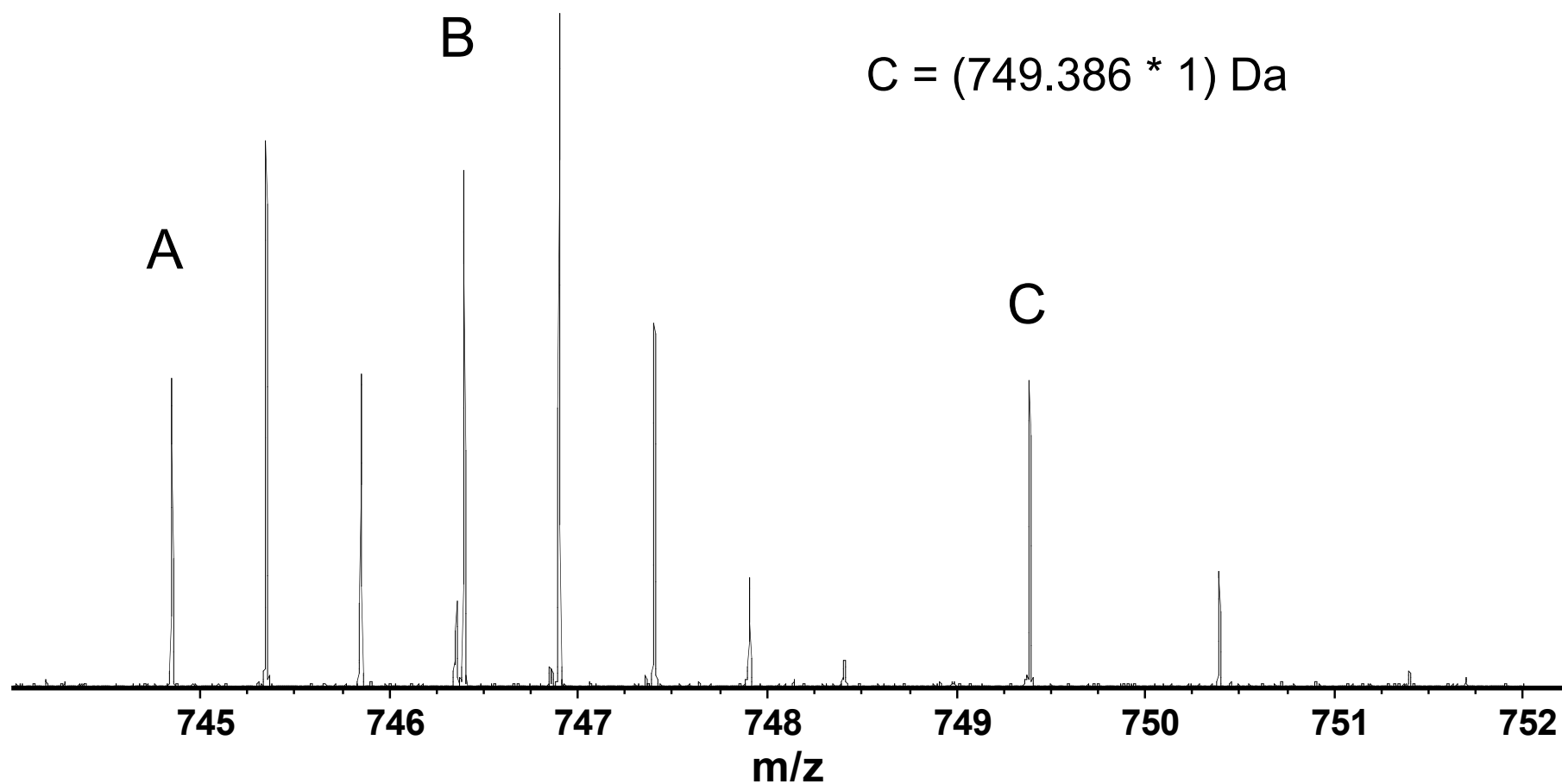
## Expanded View of 744-752 m/z Region

---

$$A = (744.842 * 2) \text{ Da}$$

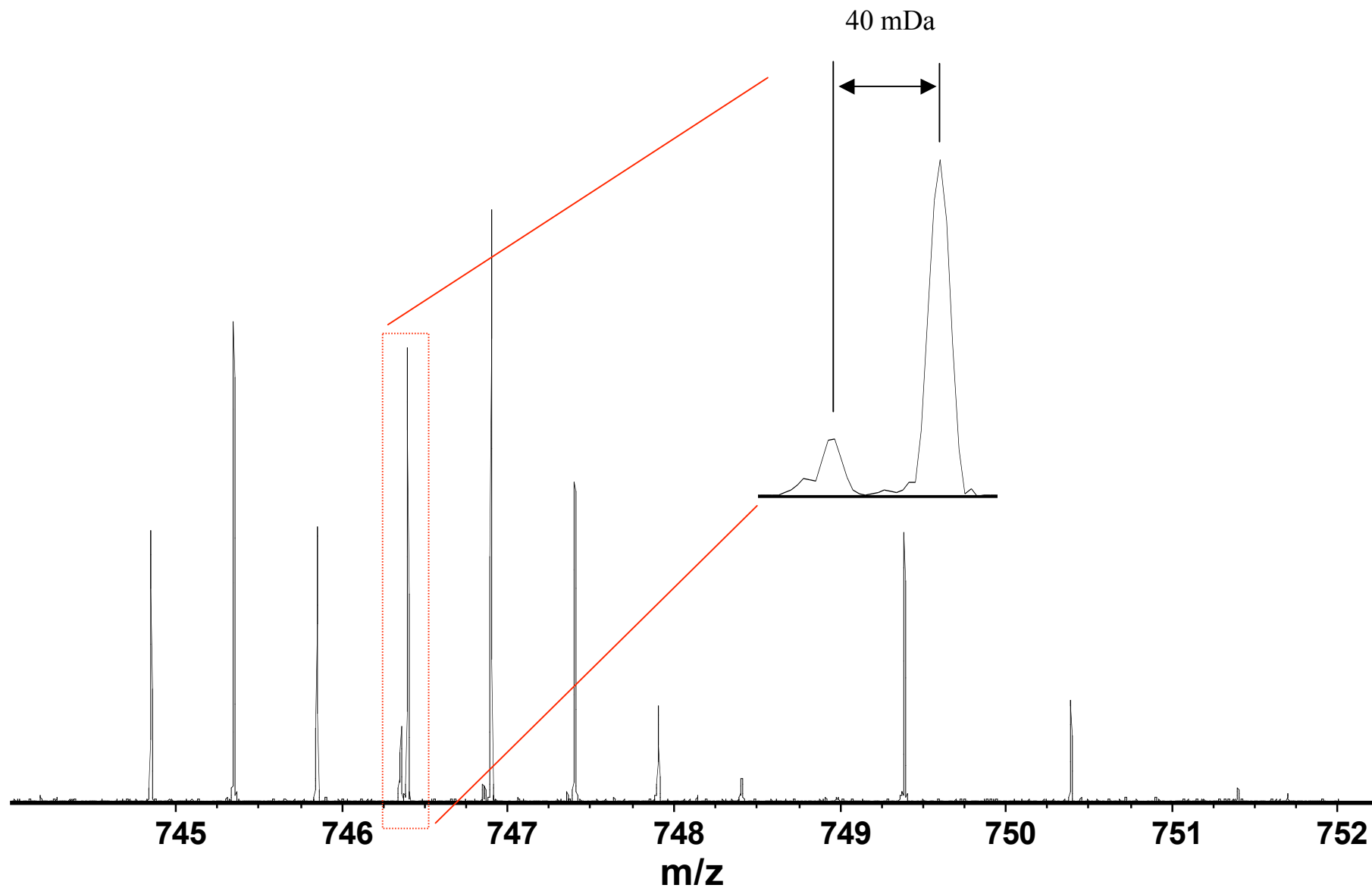
$$B = (746.390 * 2) \text{ Da}$$

$$C = (749.386 * 1) \text{ Da}$$



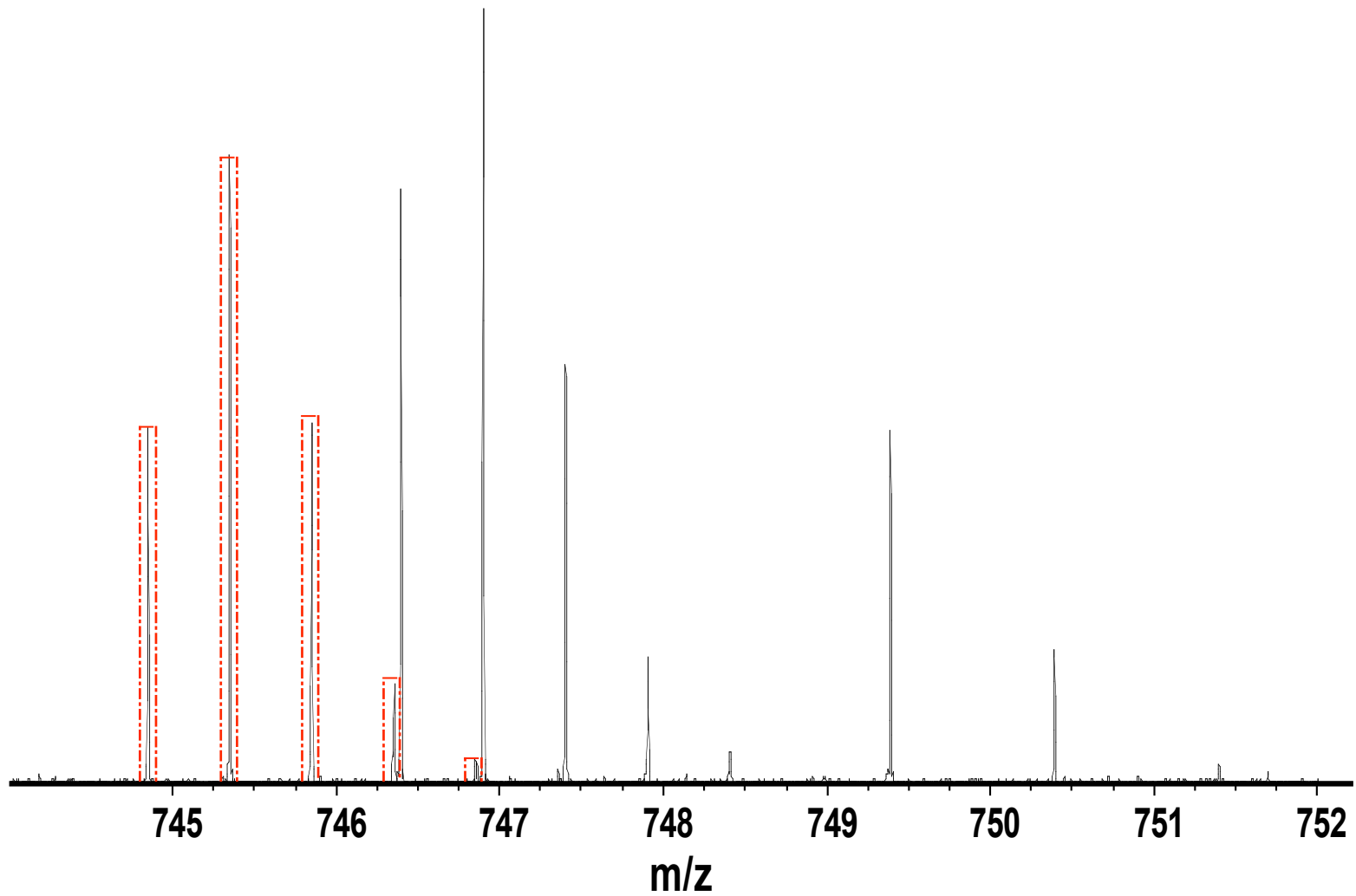
# Extremely High Resolution is Achievable with FT-MS

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# The Related Peaks are Analyzed to Determine the Distribution

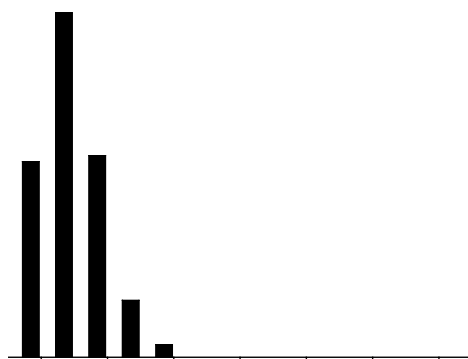
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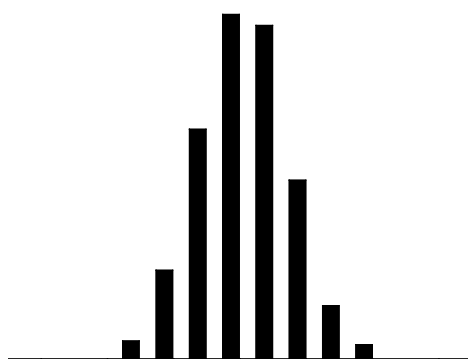
# Isotopic distributions during H/D exchange

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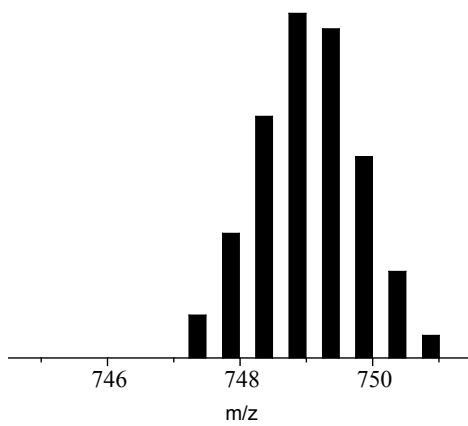
0 min



0.5 min



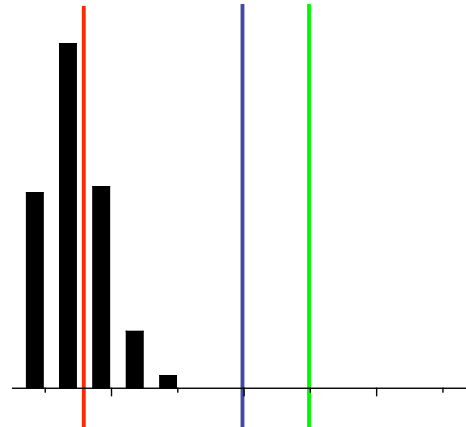
2 h



# The Centroids of the Distribution are Calculated

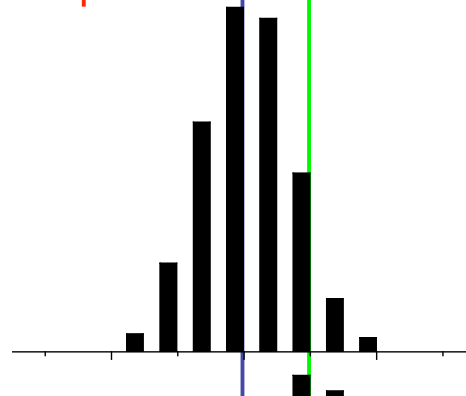
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0 min



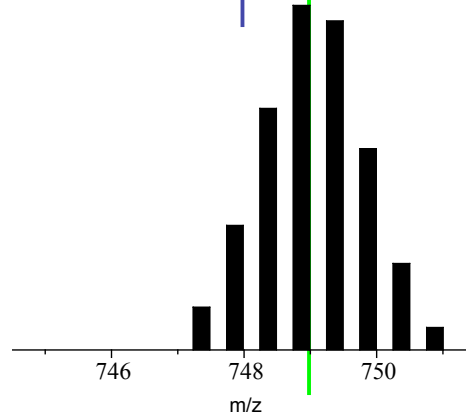
Centroid = 745.5

0.5 min



Centroid = 748

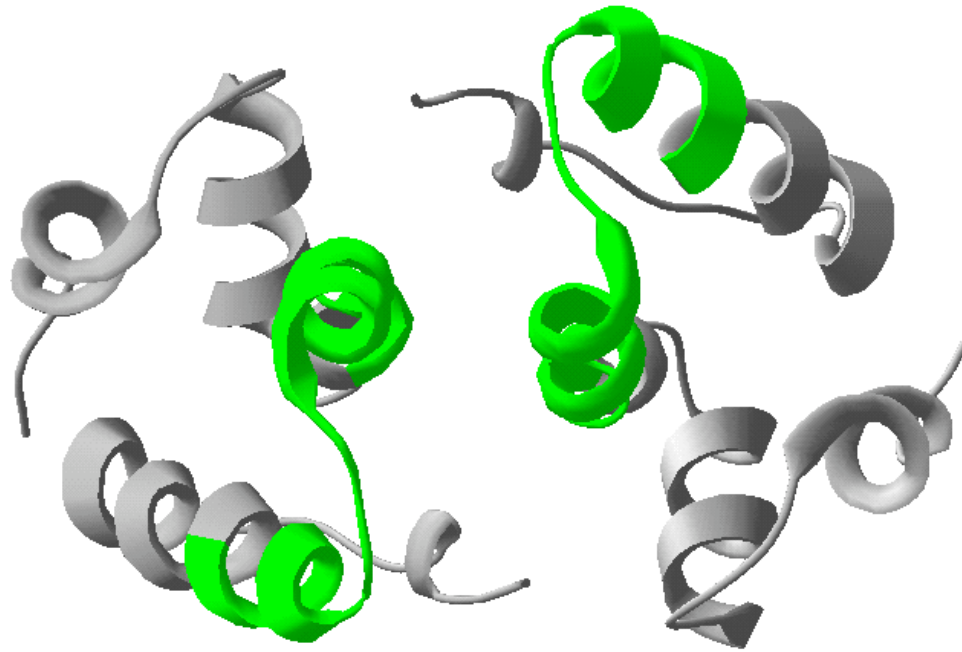
2 h



Centroid = 749

## Deuterium Incorporation at the Dimer Interface

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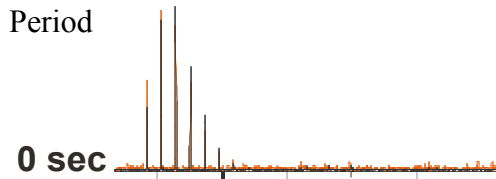




# Deuterium Incorporation Causes a Mass Increase

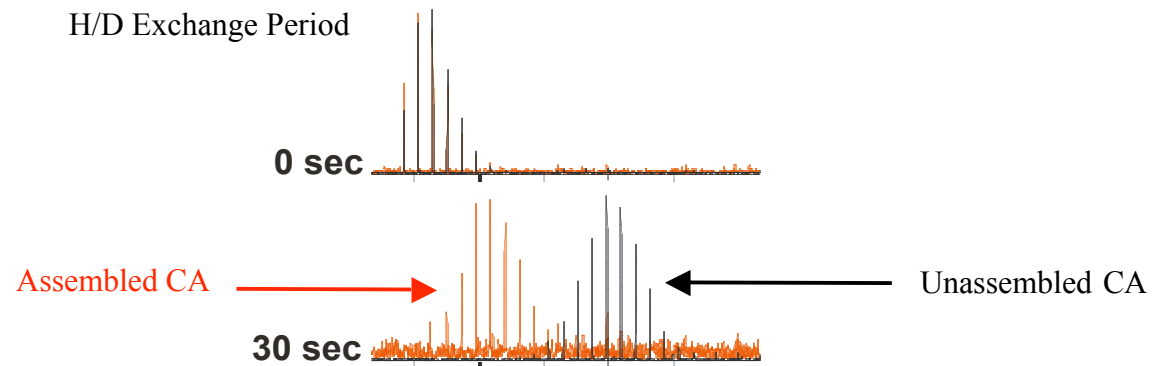
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H/D Exchange Period



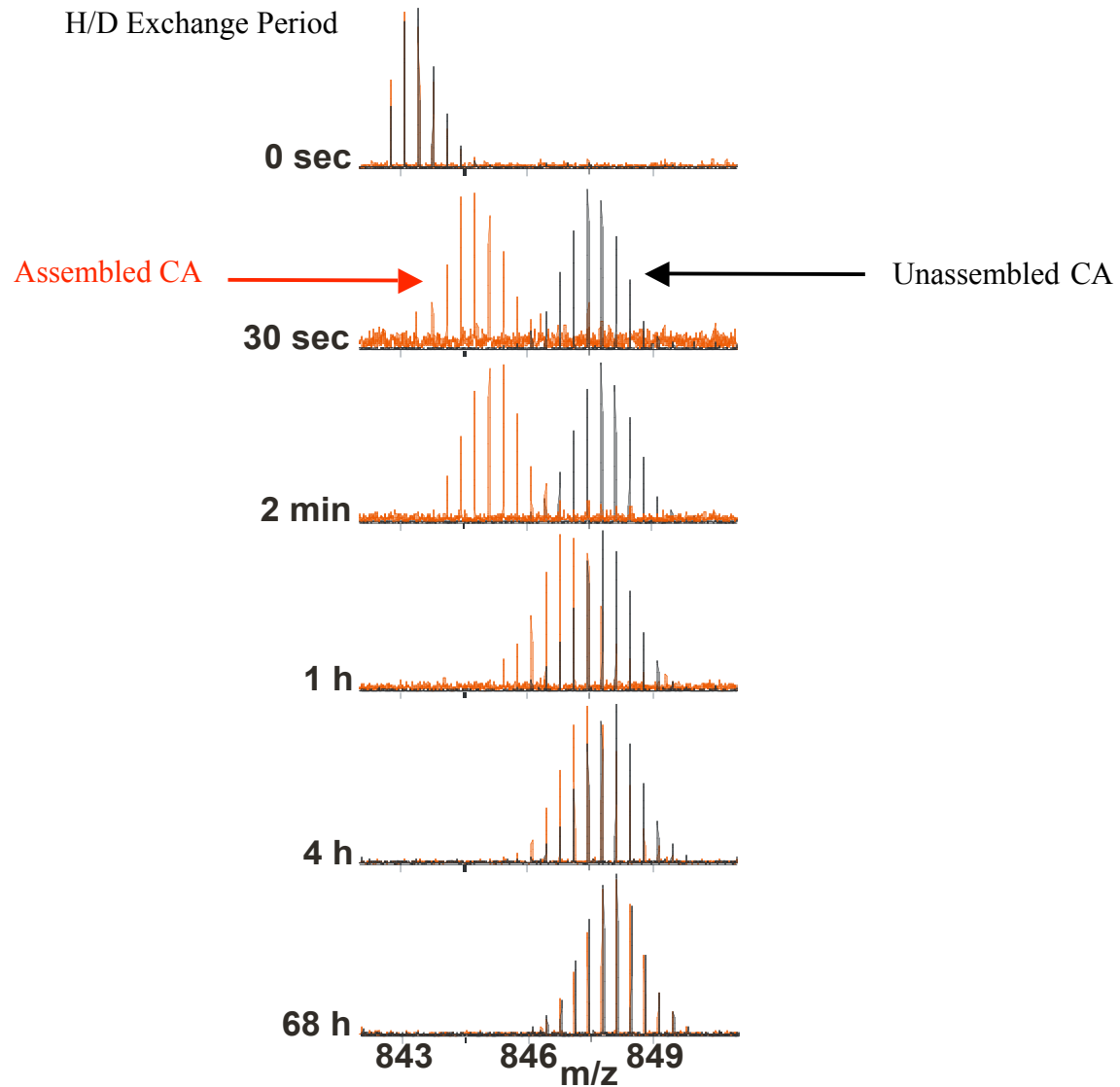
# Deuterium Incorporation Causes a Mass Increase

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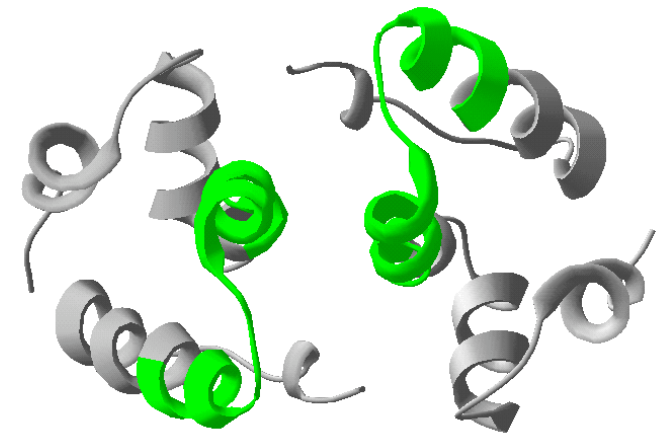
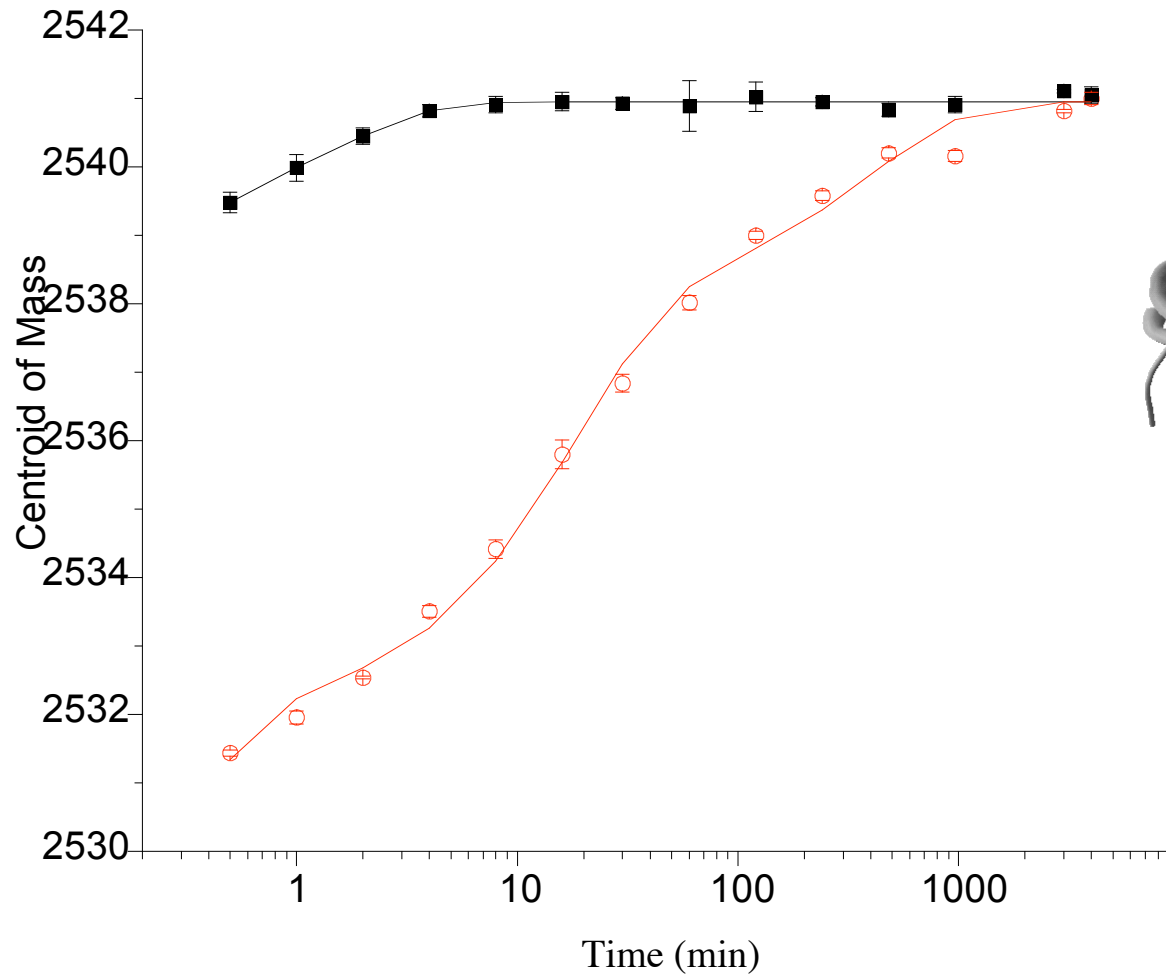
# Deuterium Incorporation Causes a Mass Increase

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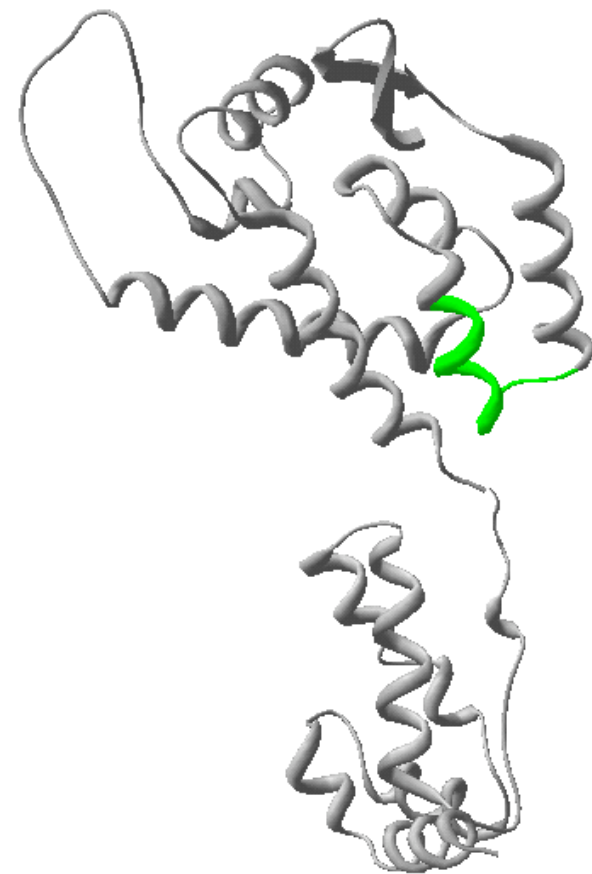
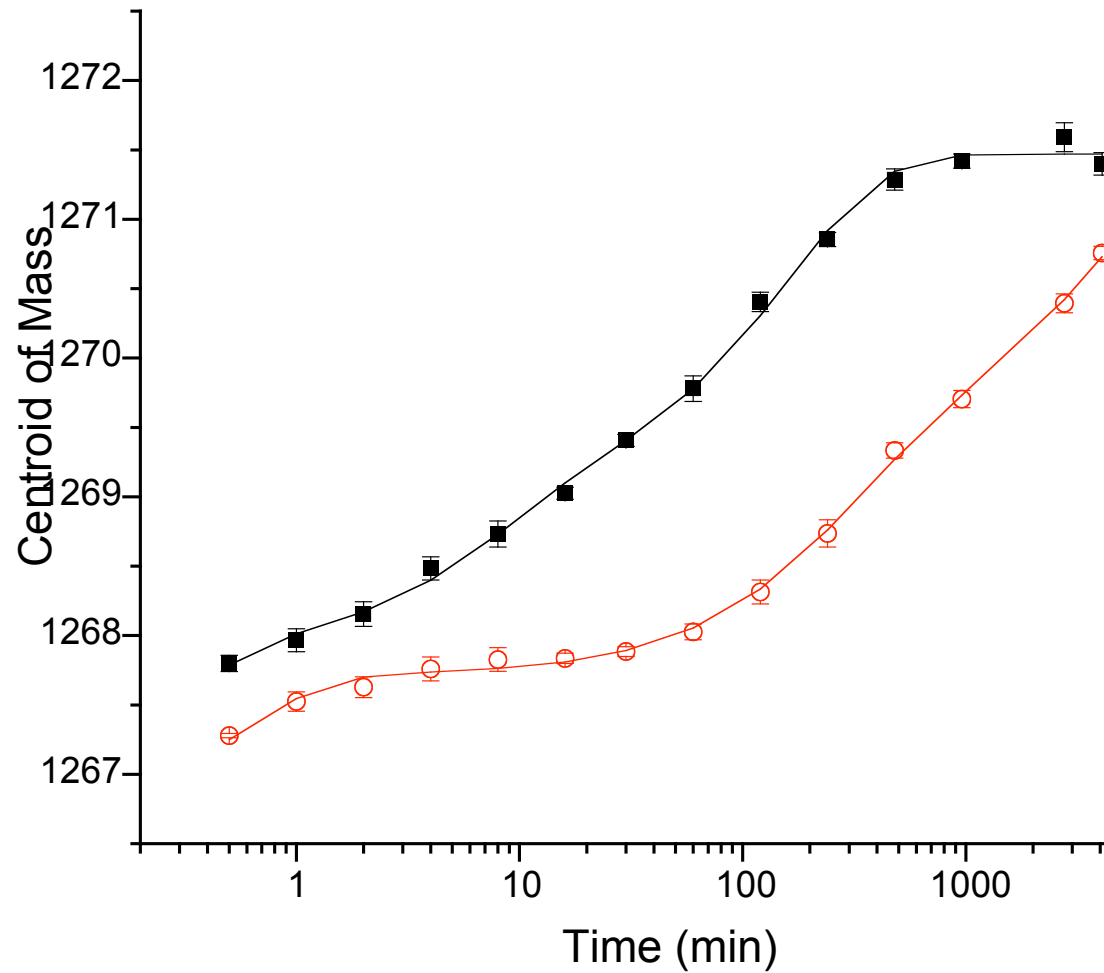
# The Dimer Interface is Protected Upon Assembly

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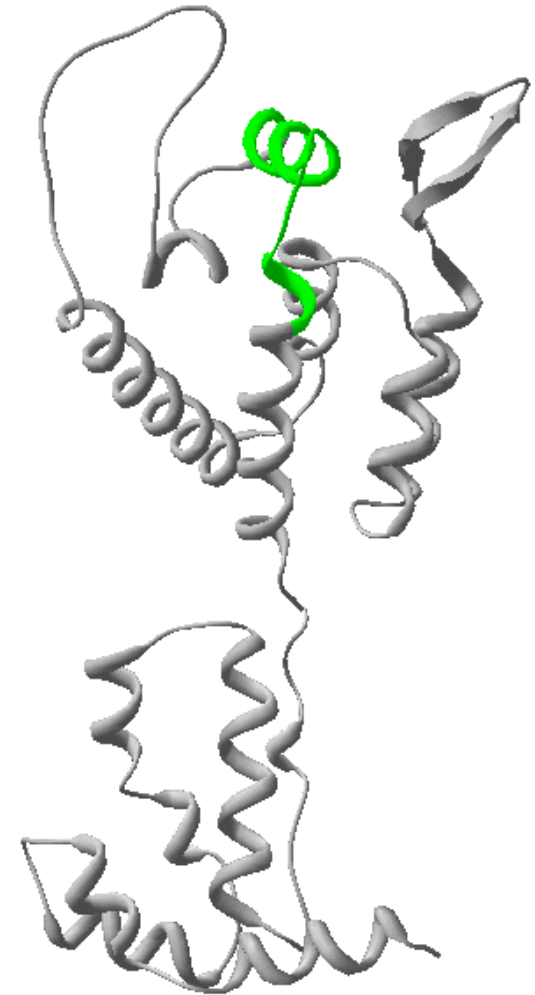
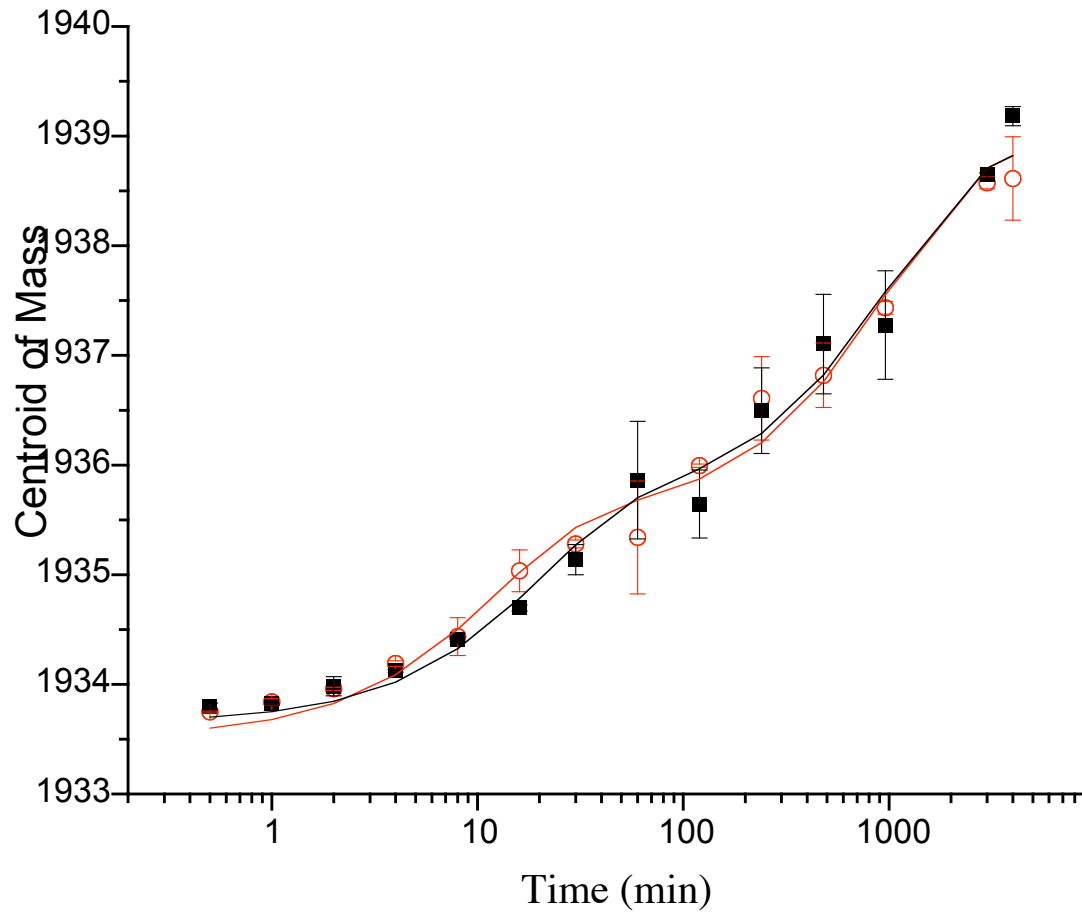
# Helix II Shows Protection Upon Assembly

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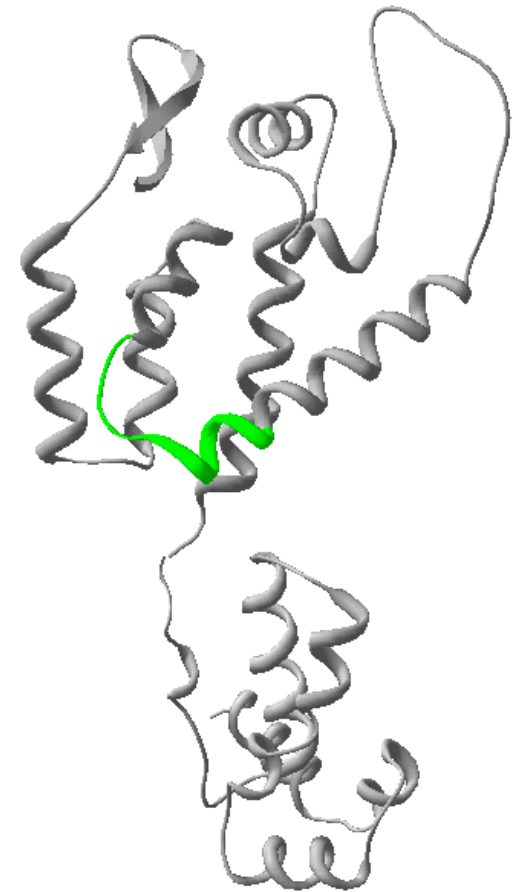
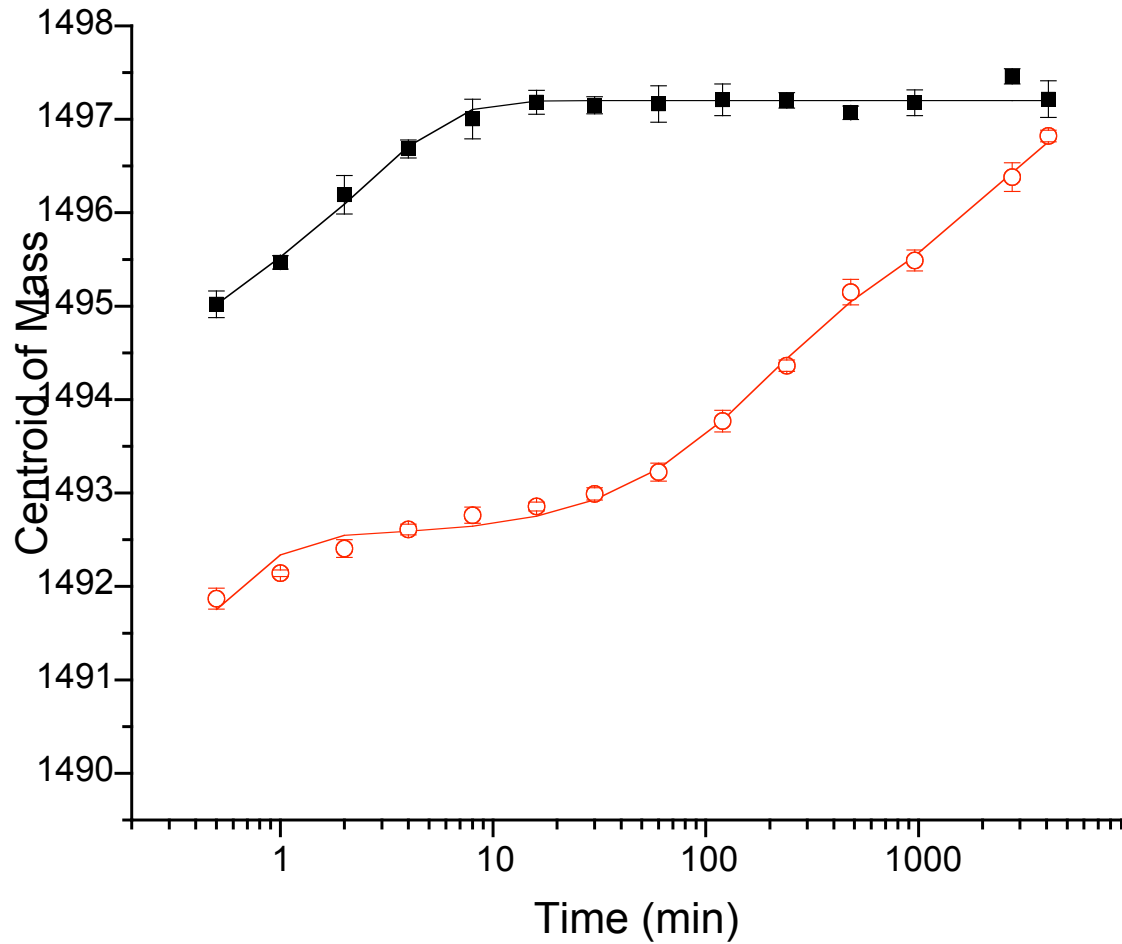


# Helices VI and VII Show Little Protection

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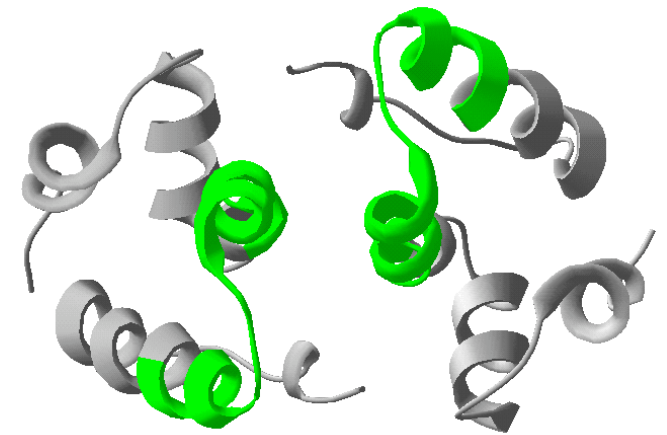
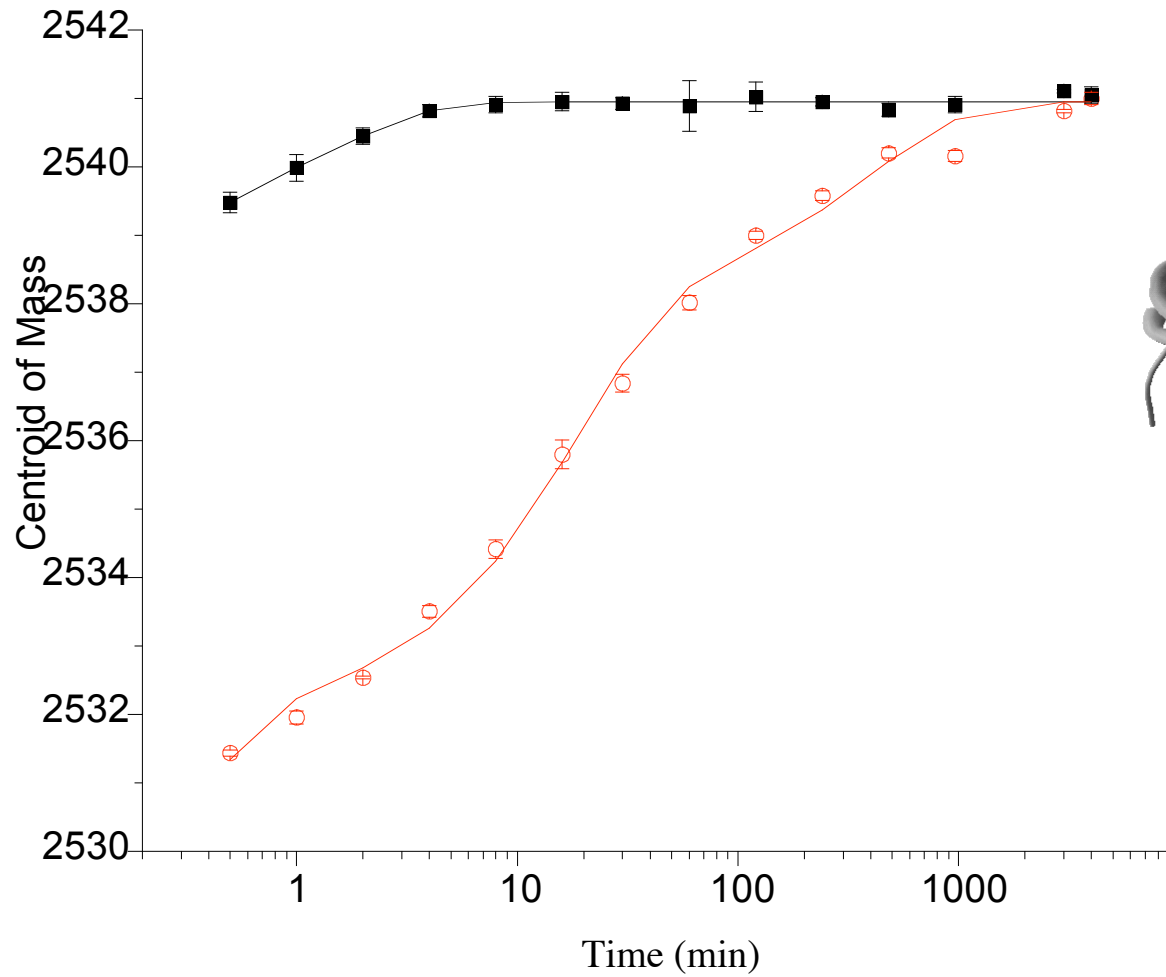


# The Bottom of Helix III becomes Protected



# The Dimer Interface is Protected Upon Assembly

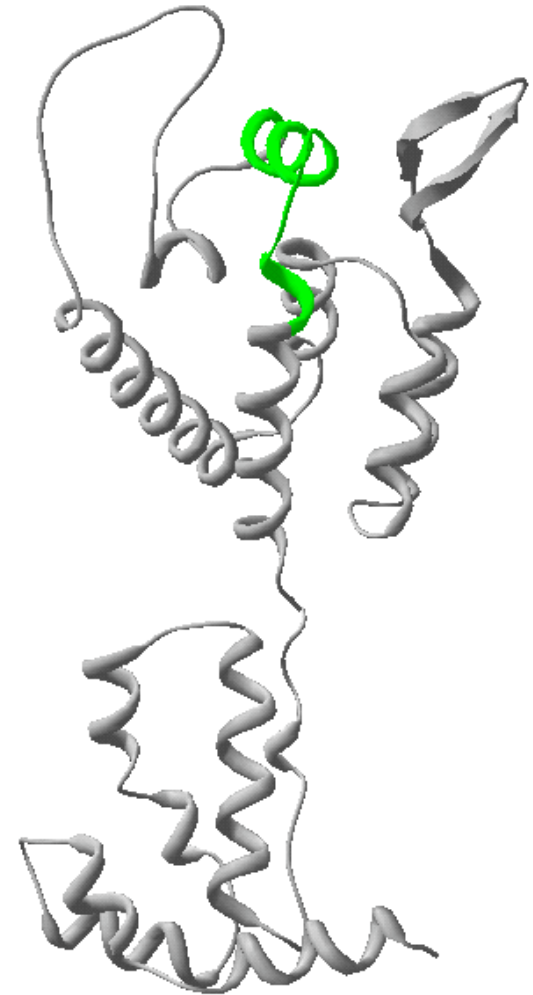
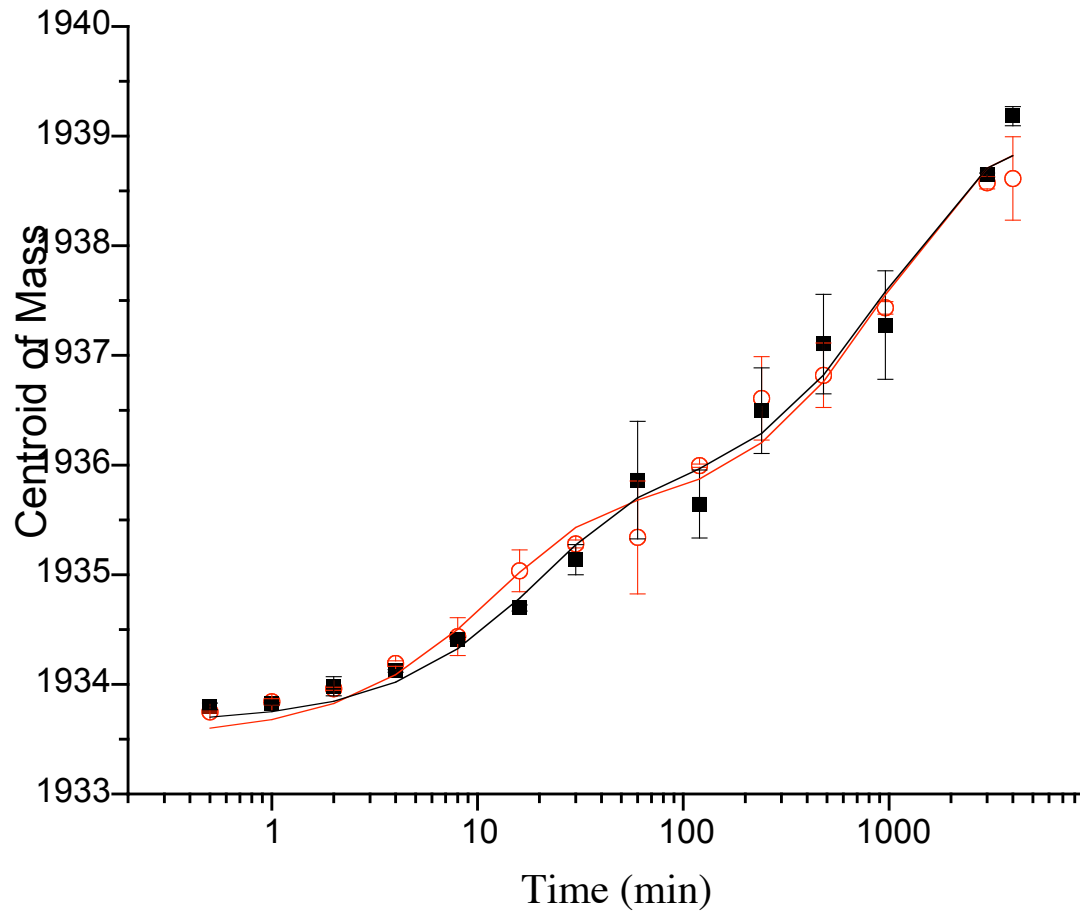
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# Helices VI and VII Show Little Protection

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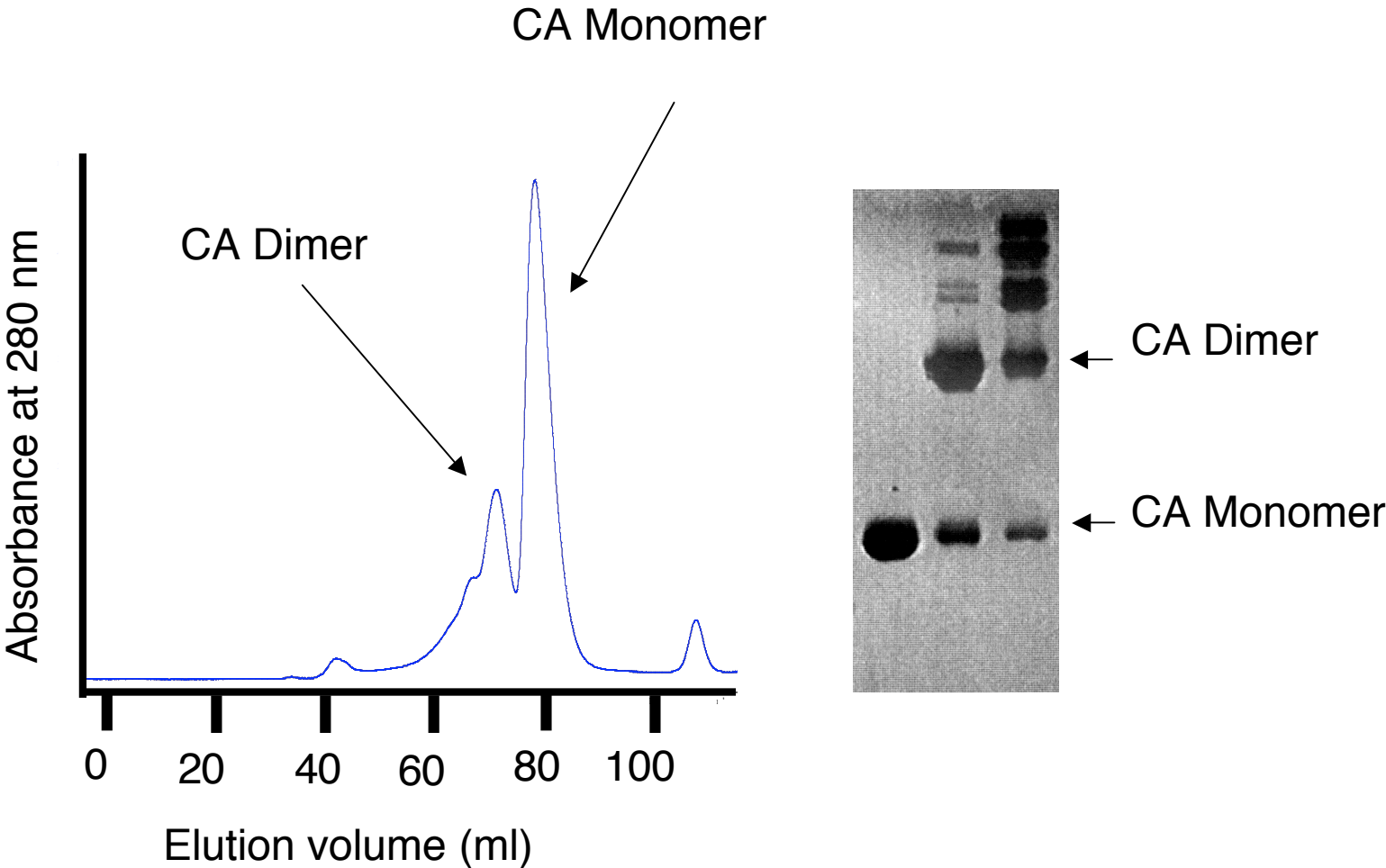


## Changes in Exchange Rates due to CA Assembly

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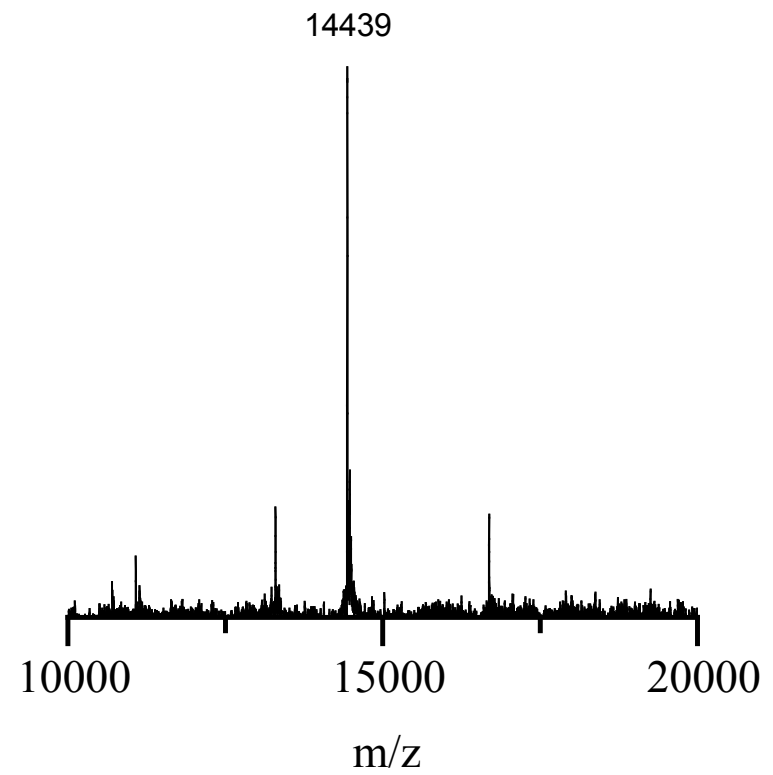
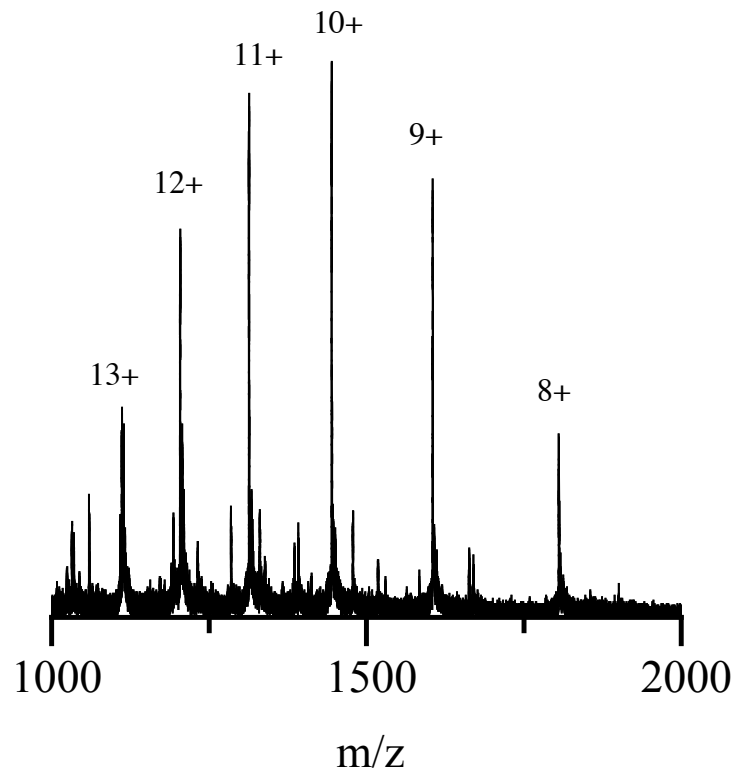


# SEC separation of cross-linked CA monomer and dimer



# Mass spectra of the cross-linked species

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## Lysine 70 was cross-linked to Lysine 182

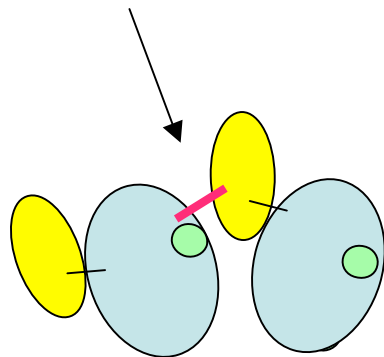
31 to 131 = 10948.9  
171 to 199 = 3375.9  
+DST = 114.0

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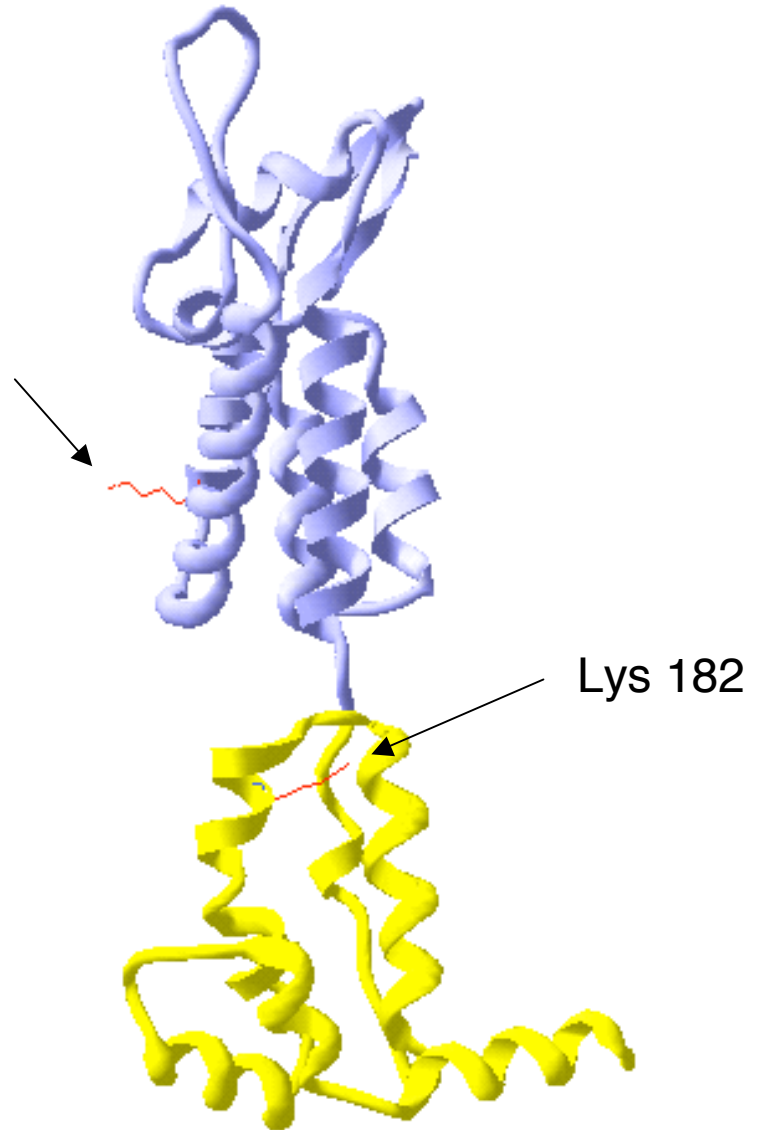
**Expected mass = 14438.8**

**Observed mass = 14439**

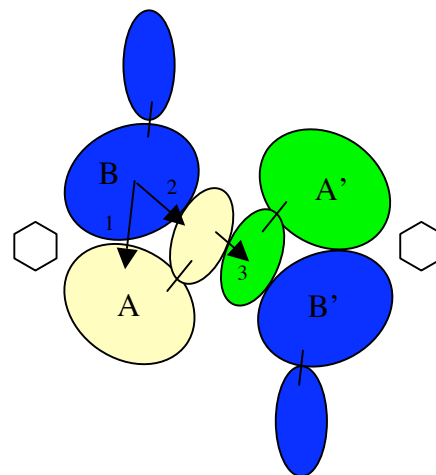
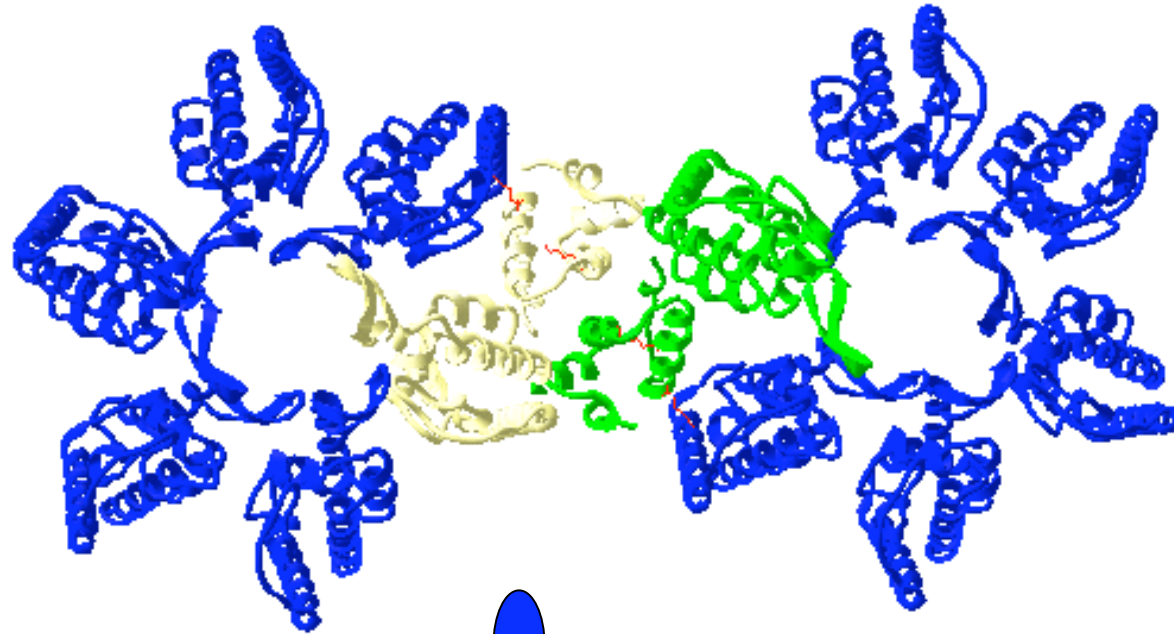
Lys70 cross-linked to 182



Lys 70

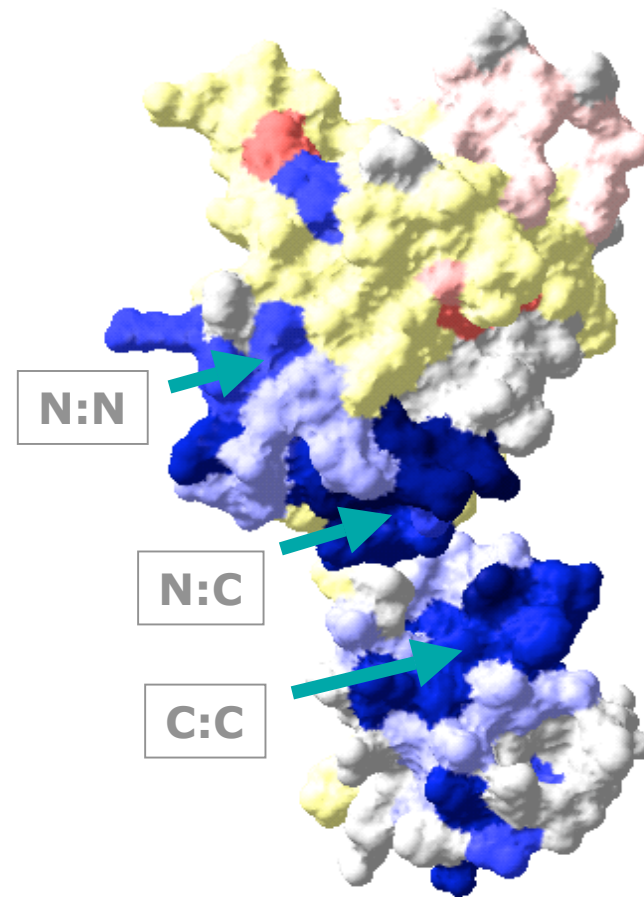


# An N-domain to C-domain Interaction in CA Assembly



# Three Sites of Interaction During Assembly

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# Advantages of Hydrogen/Deuterium Exchange

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- Small quantities required ( $10^{-12}$  mole)
- Needn't be pure
- No symmetry constraints
- Can provide time resolved or dynamic information



# The future of H/D exchange

Multi protein macromolecular complexes

Cooperativity in large proteins or macromolecular complexes

Exchange rates for individual amide protons

Protein dynamics during motor motions